B cell-targeting chimeric antigen receptor T cells as an emerging therapy in neuroimmunological diseases



Aiden Haqhikia, Georg Schett, Dimitrios Mougiakakos

oa

Summary

Background Neuroimmunology research and development has been marked by substantial advances, particularly in the treatment of neuroimmunological diseases, such as multiple sclerosis, myasthenia gravis, neuromyelitis optica spectrum disorders, and myelin oligodendrocyte glycoprotein antibody disease. With more than 20 drugs approved for multiple sclerosis alone, treatment has become more personalised. The approval of disease-modifying therapies, particularly those targeting B cells, has highlighted the role of immunotherapeutic interventions in the management of these diseases. Despite these successes, challenges remain, particularly for patients who do not respond to conventional therapies, underscoring the need for innovative approaches.

Recent developments The approval of monoclonal antibodies, such as ocrelizumab and ofatumumab, which target CD20, and inebilizumab, which targets CD19, for the treatment of various neuroimmunological diseases reflects progress in the understanding and management of B-cell activity. However, the limitations of these therapies in halting disease progression or activity in patients with multiple sclerosis or neuromyelitis optica spectrum disorders have prompted the exploration of cell-based therapies, particularly chimeric antigen receptor (CAR) T cells. Initially successful in the treatment of B cell-derived malignancies, CAR T cells offer a novel therapeutic mechanism by directly targeting and eliminating B cells, potentially overcoming the shortcomings of antibody-mediated B cell depletion.

Where next? The use of CAR T cells in autoimmune diseases and B cell-driven neuroimmunological diseases shows promise as a targeted and durable option. CAR T cells act autonomously, penetrating deep tissue and effectively depleting B cells, especially in the CNS. Although the therapeutic potential of CAR T cells is substantial, their application faces hurdles such as complex logistics and management of therapy-associated toxic effects. Ongoing and upcoming clinical trials will be crucial in determining the safety, efficacy, and applicability of CAR T cells. As research progresses, CAR T cell therapy has the potential to transform treatment for patients with neuroimmunological diseases. It could offer extended periods of remission and a new standard in the management of autoimmune and neuroimmunological disorders.

Introduction

Neuroimmunology has witnessed major advances in immunotherapeutic approaches, with various drugs targeting a range of cellular and subcellular immunological processes. In multiple sclerosis alone, more than 20 approved drugs are available, allowing some personalisation of treatment on the basis of disease activity, patient preference, and tolerability. In less common neuroimmunological disorders, such as myasthenia gravis, neuromyelitis optica spectrum disorders, and myelin oligodendrocyte glycoprotein antibody disease, disease-modifying therapies, including B-cell-targeting drugs, have also been approved or are in late-stage trials.

Monoclonal antibodies that target B cells have an increasingly important role. Rituximab has been in offlabel use for various neuroimmunological disorders for around two decades. Ocrelizumab, approved by the US Food and Drug Administration (FDA) and the European Medicines Agency (EMA) for relapsing and primary progressive multiple sclerosis, and ofatumumab, approved by the US FDA and the EMA for relapsing multiple sclerosis, are directed against CD20 and have been effective in reducing disease activity while displaying an acceptable safety profile.¹ Additionally, the recent approval of inebilizumab,² an anti-CD19 antibody,

for treatment of neuromyelitis optica spectrum disorders reflects the ongoing efforts to address B-cell activity in neuroimmunological disorders. However, some individuals with neuroimmunological disorders, such as myasthenia gravis or neuromyelitis optica spectrum disorders, do not have reduced disease activity with antibody-mediated B-cell depletion, and there is growing evidence that, despite suppressing relapses in multiple sclerosis, disease progression continues.³

In this Rapid Review, we provide an overview of cell-based therapies, particularly B cell-targeting chimeric antigen receptor (CAR) T cells, and explore their potential to transform the treatment of neuroimmunological disorders. We take into account the swiftly moving translational research that allows the interdisciplinary use of CAR T cell therapies in neurology and neuroimmunology, and take stock of past experience from the field of haemato-oncology. We also discuss the latest developments and planned CAR T trials in neuroimmunological disorders.

CART cells targeting B cell-derived malignancies Therapeutic principle and lessons learned from treating cancer patients

CAR T cells have revolutionised the treatment of B cellderived neoplasms, such as B-cell lymphoma and

Lancet Neurol 2024; 23: 615-24

Department of Neurology, Otto-von-Guericke University. Magdeburg, Germany (Prof A Haghikia MD); German Center for Neurodegenerative Diseases (DZNE), Magdeburg, Germany (Prof A Haghikia); Department of Internal Medicine 3-Rheumatology and Immunology and Deutsches Zentrum Immuntherapie (DZI). Friedrich Alexander Universität Erlangen-Nürnberg and Universitätsklinikum Erlangen. Erlangen, Germany (Prof G Schett MD); Department of Haematology, Oncology, and Cell Therapy and Oncology and Health Campus Immunology, Infectiology, and Inflammation (GCI3), Otto-von-Guericke University. Magdeburg, Germany (Prof D Mougiakakos MD)

Correspondence to: Prof Aiden Haghikia, Department of Neurology, Otto-von-Guericke University, 39120 Magdeburg, Germany

aiden.haghikia@med.ovgu.de

or

Prof Dimitrios Mougiakakos, Department of Haematology and Oncology, Otto-von-Guericke University, 39120 Magdeburg, Germany dimitrios.mougiakakos@med. ovgu.de

For more on approved drugs for multiple sclerosis see https://www.nationalmssociety.org/ Treating-MS/Medications leukaemia.4 CAR T cells are created by genetically engineering T cells to target other cells by identifying specific cell surface antigens. CARs are artificial receptors and consist of an antibody fragment that functions as the antigen-binding domain, a hinge region, a transmembrane domain, and one or more intracellular signalling domains (figure 1A).5 The antigen-binding domain, which in most cases is a singlechain variable fragment, enables binding to the target antigen without the need for it to be presented by MHC. The hinge region connects the antigen-binding domain with the rest of the receptor, and the transmembrane domain anchors the CAR into the T cell's membrane. The intracellular part of the CAR contains the parts of the T-cell receptor (TCR) that initiate T-cell activation upon antigen recognition. From the second generation of constructs onwards, CARs also incorporate one or more co-stimulatory domains (eg, CD28) to enhance T-cell activation, proliferation, and survival. After binding their antigen, CAR T cells proliferate and release cytotoxic molecules that kill their target cell (figure 1B, C). Each individual CAR T cell can destroy multiple cells. Clinically approved CAR T cells are indicated exclusively for B-cell neoplasms, including lymphomas, leukaemias, and multiple myeloma. The most widely used constructs are directed against CD19, which can be found on B cells (from the differentiation state of pro-B cells to plasmablasts; figure 1C),6 and a number of B cell-derived malignancies.7 Approved products are acquired through apheresis of autologous lymphocytes (figure 1B). Some cell separation strategies already exist, where CD4 and CD8 CAR T cells are reinfused in a fixed ratio after CD4 and CD8 T cells are separated and processed to limit toxicity.8 However, persistent CAR T cells found in longterm survivors also display a CD4 phenotype.9 The use of autologous preparations might negatively

production of CAR T cells, because the fitness of T cells can be compromised by cytotoxic (or immunosuppressive) therapies or the underlying disease.¹⁰ Strategies to overcome this issue include, among others, the use of allogeneic CAR T cells.11 Upon collection, cells are stimulated in vitro to facilitate genetic modification leading to CAR expression, which is done by gene transfer with a viral vector. Manufacturing of CAR T cells can last for up to 4 weeks. At present, rapid protocols, such those using T cells with stem cell characteristics, are being investigated.¹² After their expansion, CAR T cells are reinfused following a process known as lymphodepletion. In most cases, the chemotherapeutics cyclophosphamide and fludarabine are administered for lymphodepletion, which promotes proliferation and potential activation of the infused CAR T cells.13

Despite the success of CAR T-cell therapy, clinically significant therapy-associated toxic effects have been observed, some of which can be life-threatening. These toxic effects include cytokine release syndrome (CRS) and immune effector cell-associated neurotoxicity syndrome (ICANS).14 During CRS, activated CAR T communicate with myeloid cells, which release large amounts of inflammatory mediators, leading to sepsis-like symptoms, such as fever or hypotension. Standard therapy for CRS is IL-6 blockade, because myeloid cell-derived IL-6 has a crucial role in triggering symptoms. ICANS occurs in 20-70% of patients treated with anti-CD19 CAR T cells and varies both in terms of severity and quantity of symptoms, but typically results in a toxic encephalopathy.¹⁵ Early symptoms of ICANS include dysgraphia, wordfinding difficulties, tremor, cognitive impairment, and fatigue, which require consistent monitoring. In more severe cases, epileptic seizures, increased intracranial pressure, and even coma can occur. Although the mechanisms of ICANS remain unclear, disruption of the

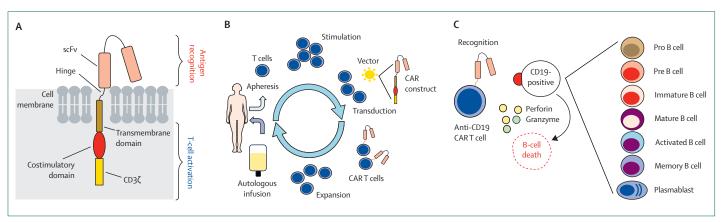


Figure 1: Design, production, and mode of action of CART cells

(A) A prototypical second-generation CAR with an antibody-derived antigen-binding single-chain variable fragment (scFv), hinge region, transmembrane domain, costimulatory domain (eg, CD28 or 4-1BB), and CD3ζ-chain of the T cell receptor. (B) To produce CART cells, patients undergo lymphapheresis. Upon collection, autologous T cells are stimulated in vitro (eg, by triggering the T cell receptor or co-stimulatory molecules, such as CD28) and genetically modified by, for example, transduction with viral vectors to express a CAR. The CART cells produced in this way will be further expanded and subsequently reinfused into the patient after lymphodepletion (done with a combination of fludarabine and cyclophosphamide). The depicted process typically requires up to 4 weeks. (C) CART cells, directed, for example, against CD19, recognise CD19 expressed at various stages of the B-cell lineage, become activated, and destroy the target cell by releasing effector molecules, such as granzyme and perforin. CAR=chimeric antigen receptor.

blood-brain barrier as a result of activated endothelial cells has a role in its pathophysiology.¹⁶ Due to the permeable blood-brain barrier, inflammatory mediators can enter the CNS causing neuronal dysfunction. Unlike CRS, IL-6 blockade is ineffective in other neurotoxicities, and steroids are mainly used.17 Risk factors for CRS or neurotoxicity comprise high tumour burden, systemic inflammation (eg, elevated CRP or ferritin), and preexisting neurological conditions predisposing to neurotoxic effects.¹⁸ Cytopenia is another adverse effect that, depending on timing and duration (ie, <3 vs >3 months), is linked to haematotoxic lymphodepletion or immunological processes.¹⁹ Extended cytopenia can require a stem cell boost, leaving patients susceptible to infectious complications. As anticipated, anti-CD19 CAR T cells also eradicate non-malignant B cells (on-target, offtumour effect), resulting in prolonged B-cell depletion, sometimes necessitating a year-long therapy for immunoglobulin replacement.14

CART cells in patients with autoimmunopathies

The prolonged and deep depletion of B cells in cancer patients receiving CAR T-cell therapy was used as the rationale for their first use in rheumatoid autoimmune diseases.20 Unlike B cell-targeting antibodies, such as rituximab, CAR T cells are autonomous and do not require natural killer (NK) cells, macrophages, or the complement system²¹ to perform their function, even deep within tissues. This autonomous mechanism might be the reason why the deployment of therapeutic antibodies often does not result in the same degree of B-cell depletion as seen in patients treated with CAR T cells, especially in compartments of interest, such as the CNS (table 1).22 In autoimmune diseases, B cells not only act as producers of autoantibodies, but they can also present self-peptides through MHC, thereby activating autoreactive T cells.23 Several reports, including those related to systemic lupus erythematosus, 24 antisynthetase syndrome, 25 and scleroderma, 26 indicate that anti-CD19 CAR T cells could generate long-lasting remissions in therapy-resistant cases. In a German case series, patients with treatment-refractory severe forms of systemic lupus erythematosus, myositis, or scleroderma remained disease-free during a median follow-up of 15 months (range 4-29) and despite reappearance of B cells in 14 of 15 patients a mean of 112 days (SD 50) after infusion.27 These findings could affect the treatment of autoimmune diseases, and several early phase clinical trials evaluating B cell-directed CAR T cells have been initiated or are planned (>10 studies in systemic lupus erythematosus).28 Although the patient population is small, available case series present compelling discoveries-ie, despite infusion of CAR T cells into a pro-inflammatory or autoimmune context, no augmented toxicity signals have been observed.20 Additionally, the quantity of targeted CD19-positive cells (ie, B-cells) is substantially lower than in B cell-derived

	Monoclonal antibodies	CART cells
Availability	Prompt, off the shelf	Around 4 weeks, individual production*
Persistence	Limited	In-vivo expansion
Biodistribution	Slow, passive	Fast, passive and migration
Mode of action	Complement-dependent cytotoxicity, cell-mediated cytotoxicity, cell-mediated phagocytosis	Release of cytotoxic molecules
CAR=chimeric antig	gen receptor. *Allogeneic CART cells are in development.	
Table 1: Therapeu	tic differences between monoclonal antibodies and	CART cells targeting B cells

malignancies, resulting in less CAR T-cell mediated toxicity.¹⁸ Despite the substantial reduction or even complete absence of autoantibody production following infusion of CAR T cells, most patients with rheumatic conditions maintain sufficient immunoglobulin concentrations, and their protective vaccination titres are maintained.²⁰

An explanation of this paradox might be that CD19 is still present on plasmablasts but is no longer on long-lived plasma cells.²⁹ This explanation would lead to the following hypotheses: (1) CD19-postitive, CD20-negative plasmablasts are central to autoimmunity; and (2) protective antibodies are still produced in sufficient amounts by CD19-negative, CD20-negative, long-lived plasma cells (in the bone marrow). The compelling efficacy of CAR T cells in the treatment of rheumatoid autoimmune diseases, together with the favourable safety data, provide the impetus to pursue such strategies in B cell-driven neuroimmunological disorders.

Role of B-cells in neuroimmunological disorders

Antibody-mediated depletion of B cells (ie, by anti-CD19 or anti-CD20 therapeutic antibodies) does not necessarily lead to clinical stabilisation of neuroimmunological disorders.3,30,31 One possible explanation could be the prevalence of TBX21-high (or T-bet-high) memory B-cells, which drive chronic inflammation. They do not circulate but reside in the tissue close to the site of inflammation and can adopt a double-negative (CD19-negative, CD20-negative) phenotype.³² Additionally, in the context of autoimmunity, B cells have shown to be producers of pro-inflammatory cytokines, such as IFNy, IL-6, and GM-CSF, as well as autoantibodies.^{33,34} This feature allows cytokine-producing B cells to drive and to maintain the formation of tertiary lymphoid structures, which could cause disease progression in multiple sclerosis despite anti-inflammatory interventions and disease-modifying therapy. As shown extensively in multiple sclerosis, for which a specific autoantibody has not been identified, inhibition of B-cell functions, such as cytokine production, antigen presentation, and chronic tissue inflammation, might be the main treatment effect of B-cell depletion.35 Therefore, the ability of CAR T cells to penetrate into the tissue and sufficiently deplete these otherwise inaccessible B cells that drive chronic, tissue-resident inflammation

could explain the rapid and long-lasting therapeutic effect of CAR T cells.

Myasthenia gravis

Myasthenia gravis is an antibody-mediated neuroimmunological disorders and the most common neuromuscular disease. About 90% of the seropositive cases have antibodies against the acetylcholine receptor (AChR), and the remaining myasthenia gravis types (eg, muscle-specific tyrosine kinase [MuSK] antibody-positive cases or other autoimmune neuromuscular disorders) are much rarer and can be associated with malignancies.36 Although the main pathological mechanism is autoantibody-mediated blockade of AChR, complement activation leads to destruction of the receptor and renders the muscle unresponsive to nerve signals. Several studies (case series and open-label trials) have investigated the therapeutic efficacy of rituximab. Although patients with anti-MuSK antibody-positive myasthenia gravis responded favourably to B-cell depletion, rituximab did not result in stabilisation for all patients with anti-AChR-positive myasthenia gravis.37 This observation has led to the introduction of complement-targeting antibodies, such as eculizumab38 and ravulizumab,39 or the overall reduction of immunoglobulins by efgartigimod.40 These therapeutic approaches have proven effective in highly active myasthenia gravis. However, their use is associated with increased infections, high frequency of administration, or (sometimes prolonged) hypogammaglobulinaemia.

B cell-targeting CAR T cells might lead to a sustained suppression of disease activity. A US phase 1b/2a study41 investigated the safety of RNA-engineered CAR T cells targeting B-cell maturation antigen (BCMA) in 14 adults with myasthenia gravis and suggested that the therapy is safe to use. In a 12-month update of the study, 42 five of the seven patients treated continued to show clinical improvement in MG-ADL, QMG, and MGC scores. In a 33-year-old woman with highly active myasthenia gravis (despite previous treatment with rituximab, bortezomib, and mycophenolate), anti-CD19 CAR T cells led to longterm disease stabilisation with a good safety profile.43 Overall, IgG concentrations remained stable, and no increased susceptibility to infections was observed. In summary, available data are encouraging, with the forthcoming clinical trials (table 2) expected to yield further insight into the potential of anti-CD19 CAR T cell therapy as a durable treatment option for myasthenia

Antibody-mediated neuroimmunological disorders

Progress in diagnostics, including MRI and serum and CSF biomarkers, has allowed for a clearer differentiation of antibody-mediated neuroimmunological disorders from multiple sclerosis. 48,49 The pace of therapy development in multiple sclerosis has been matched by implementation of clinical trials in less common antibody-mediated neuroimmunological disorders, resulting in the approval

of multiple drugs for neuromyelitis optica spectrum disorders and myelin oligodendrocyte glycoprotein antibody disease. Although rituximab has been used offlabel to target B cells in cases where immunomodulatory therapies, such as azathioprine and mycophenolate, did not elicit a response, its efficacy is only supported by retrospective data.50 Similarly, the diverse and expanding set of antibody-mediated autoimmune encephalitides have been subject to empirical treatment. The range of immunotherapies used in this category varies from IVIG, steroids, and plasma exchange in the acute stage to rituximab and, the plasma cell-targeting drug, bortezomib in refractory cases.⁵¹ Antibodies targeting cytokines (eg. tocilizumab) or complement (eg, eculizumab) have also been used to treat autoimmune encephalitides. 52 The first drug approvals for the treatment of antibody-mediated neuroimmunological disorders have been granted for the anti-aquaporin-4-positive neuromyelitis optica spectrum disorders following randomised clinical trials that investigated the anti-CD19, B cell-depleting antibody inebilizumab,2 the complement-binding and complementneutralising antibodies eculizumab⁴⁶ and ravulizumab,⁵³ and the anti-IL6 receptor antibody satralizumab.54 These advancements offer a potent therapeutic arsenal, with safety data requiring long-term evaluation. Similar challenges to those in myasthenia gravis are anticipated. In the context of neuromyelitis optica spectrum disorders, characterised by severe inflammatory attacks, achieving long-term remission is the primary objective. An initial phase 1 trial⁵⁵ in 12 adults with neuromyelitis optica spectrum disorders using anti-BCMA CAR T cells, although not primarily assessing clinical efficacy, showed that 11 patients remained drug-free and relapse-free after a median of 5.5 months follow-up. Another trial using a tandem CAR T cell against CD19 and CD20 has been announced in China, but the results have not yet been reported (table 2). In conclusion, CAR T-cell therapy might become a valid option for severe, refractory cases of antibody-mediated neuroimmunological disorders.

Multiple sclerosis

Multiple sclerosis research has driven innovation in the field of neuroimmunological disorders. Despite absence of a specific autoantigen, many immune-mechanistic discoveries have produced a multitude of therapies that inhibit, modulate, or deplete various immune targets. Most of the investigations that led to available therapies have provided a better understanding of the intricate autoimmune and neurodegenerative nature of multiple sclerosis. B cell targeting, in particular, and its strong effect on reducing disease relapses has challenged the notion that multiple sclerosis is a primarily T cell-mediated CNS disease.56,57 The most apparent involvement of B-cells in multiple sclerosis pathophysiology are oligoclonal bands in the CSF, which are detectable in the most patients with multiple sclerosis. Three B cell-depleting antibodies, all directed against CD20, have been approved for the

Anti-CD29 Number of the control									
Anti-EOAA Warious Anti		CART cell	Disease	Study design	Number of patients	Main inclusion criteria	Primary outcomes	Status	Main results
Anti-BOMA Washenia gaois Copen label, 30 s.18 was old seropositive, and concepting and formation of manifecture, phase 2 Anti-BOMA Various and concepting a	NCT03605238	Anti-CD19/ CD20 CART cell	Neuromyelitis optica spectrum disorder	Open-label phase 1	N/A	12–75 years old, aquaporin-4-lgG seropositive, 22 relapses in the last year or 23 relapses in the last 2 years, the rapy refractory (corticosteroid plus immunosuppressant)	Study-related adverse events (12 months), annual relapse rate, EDSS, visual acuity	Announced August, 2018; estimated completion August, 2020; results not published	Withdrawn owing to difficulties in recruiting patients
Hereometric open-label, 18 18-75 years old, seropositive, Dose-limiting toxic effects Announced decoders and ordered in on- Myasthenia gravis 2 MG-ADI->6 OMG, MG-ADI stimated completion phase 1	NCT04146051	Anti-BCMA CAR T cell	Myasthenia gravis	Open label, non- randomised, multicentre, phase 2	30	≥18 years old, seropositive and seronegative (MGFA III–IV), MG-ADL>6	Safety, MG-ADL	Announced April, 2019; estimated completion December, 2023; published"	7 completed the study (7 received MTD once weekly for 6 weeks [group 2]), MG-ADL mean score change to baseline–6 (95% Cl-9 to-3) and QMG-7 (-11 to-3) after 24 weeks; group 1 (twice weekly for 3 weeks at MTD) and group 3 (once monthly for 6 months) discontinued; MTD was determined in phase 1 of the study (median 17:3×10° cells [range 9:7-33:1] divided over a median of 6 infusions [3-6])
enia gravis Myasthenia gravis 2 MG-ADL-6 QMG,MG-ADL Cause and Treatment of Inflammatory Neuropathy Inflammatory Neuropathy Inflammatory infl	61557	Anti-BCMA CAR-T cell	Various neuroimmunological disorders	Open-label, non- randomised, single-centre, phase 1	18	18-75 years old, seropositive, refract ory to standard therapy	Dose-limiting toxic effects (3 months), treatment- emergent adverse events (2 years), serum antibody titres	Announced September, 2022; estimated completion May, 2024	
inflammatory demyelinating demyelinating demyelinating bolyradiculopathy inerrotising more of a more of a myopathy in myopathy in myopathy in pectrum inflammatory into a section of a myopathy into pectrum inflammatory into a myopathy into perturbation into a myopathy into a myopath	henia gravis	:	Myasthenia gravis	:	~	MG-ADL>6	QMG, MG-ADL		Patient 1: 33-year-old woman, positive for antibodies against AChR and Titin, received 6-16×10′ cells, grade 1 CRS, improved from QMG 12 at baseline to below 5 at month 18, was seronegative for antibodies against AChR at month 18; Patient 2: 60-year-old woman, positive for antibodies against MuSK, received 5-04×10′ cells, no CRS, improved from QMG 18 at baseline to below 2 at month 18, was seronegative for antibodies against AChR at month 18**
ed Immune-mediated 1 Manual Muscle Testing score necrotising t myopathy Neuromyelitis optica 12 ≥2 relapses in the last year or Time to first relapse, EDSS, Interim analysis spectrum disorder refractory to at least one immunosuppressant for >1.9 ear	ic matory elinating diculopathy	:	Chronic inflammatory demyelinating polyradiculopathy	÷	Unknown	INCAT score of 2-9	Inflammatory Neuropathy Cause and Treatment disability score, Medical Research Council sum score	÷	Not known
Neuromyelitis optica 12 ≥2 relapses in the last year or Time to first relapse, EDSS, Interim analysis spectrum disorder ≥3 relapses in the last 2 years, visual acuity published in 2023** refractory to at least one immunosuppressant for >1 year	ne-mediated ising athy cohort		Immune-mediated necrotising myopathy	÷	₽	:	Manual Muscle Testing score	÷	25-year-old man, positive antibodies against SRP, received 6-53×10° cells, grade 1 CRS, improvement in MMT score (from 96 to 137) and negative for anti-SRP antibodies at month 18 ⁴⁵
	spectrum er cohort	:	Neuromyelitis optica spectrum disorder	i.	12	22 relapses in the last year or 23 relapses in the last 2 years, refractory to at least one immunosuppressant for >1 year	Time to first relapse, EDSS, visual acuity	Interim analysis published in 2023 ⁴⁶	12 patients completed the study; three groups were tested (dosed escalation group with two sub-groups of 3 each [0.5 x 10° cells per kg] and dose expansion group of 6 [1 x 10° cells per kg]]. 11 patients remained drug and relapse free in interim analysis after a median of 5.5 months of follow-up, 1 patient had a relapse with optic neuritis of the left eye (Table 2 continues on next page)

	CAR T cell	Disease	Study design	Number of patients	Number of Main inclusion criteria patients	Primary outcomes	Status	Main results
(Continued from previous page)	evious page)							
NCT05828225	Anti-CD19 CAR T cell	Myasthenia gravis	Open-label phase 1	6	218 years old, seropositive (AChR; MGFA IIa–IVb), MG-ADL >5, QMG >11, refractory to standard therapy	Dose-limiting toxic effects Announced April, 2023; (28 days), treatment- estimated completion emergent adverse events April, 2026 (90 days)	Announced April, 2023; estimated completion April, 2026	Notknown
NCT05451212	Anti-MuSK CAAR-T cell	MuSK myasthenia gravis	Open-label phase 1	24	≥18 years old, seropositive for MuSK and AChR (MGFA I–IVa)	Dose-limiting toxic effects, treatment-emergent adverse events (9 months)	Announced December, 2023; estimated completion October, 2028	Not known
NCT06138132	Anti-CD19 CART cell	Progressive multiple sclerosis	Open-label phase 1	12	218 years old, diagnosis according to 2017 McDonald criteria, progressive according to Lublin 2014 criteria	Dose-limiting toxic effects (12 months)	Announced December, 2023; estimated completion June, 2027	Not known
Individual case study ⁴⁷	CD19 CAR-T	CD19 CAR-T Myasthenia gravis	·	Н	218 years old, seropositive (AChR; MGFA II-IV), refractory to standard therapy	Safety, QMG, Besinger score	Published in 2023	Treatment with 1×10^{4} total cells safe, QMG change from baseline -10
Current studies testing receptor. EDSS=Expant MTD=maximum tolera	JCART cells in neu ded Disability Statı ıted dose. MuSK=n	roimmunological disorder: us Scale. INCAT=Inflammat nuscle-specific tyrosine kin	s ordered by date th tory Neuropathy Ca iase myasthenia gra	ney were register suse and Treatm avis. N/A=not av	Current studies testing CAR T cells in neuroimmunological disorders ordered by date they were registered or published. AChR-acetylcholine receptor. BCMA receptor. EDSS=Expanded Disability Status Scale. INCAT=Inflammatory Neuropathy Cause and Treatment Disability Score. MG-ADL=Myasthenia Gravis—Av MTD=maximum tolerated dose. MuSK=muscle-specific tyrosine kinase myasthenia gravis. N/A=not available. QMG=Quantitative Myasthenia Gravis score.	e receptor. BCMA=B-cell matura: thenia Gravis—Activity of Daily Li snia Gravis score.	tion antigen. CAR=chimerica iving score. MGFA=Myasthen	Current studies testing CAR T cells in neuroimmunological disorders ordered by date they were registered or published. AChR-acetylcholine receptor. BCMA=B-cell maturation antigen. CAR=chimeric antigen receptor. CAAR=chimeric autoantibody receptor. EDS5=Expanded Disability Status Scale. INCAT=Inflammatory Neuropathy Cause and Treatment Disability Score. MG-ADL=Myasthenia Gravis—Activity of Daily Living score. MGFA=Myasthenia Gravis Foundation of America (dassification). MTD=maximum tolerated dose. MuSK=muscle-specific tyrosine kinase myasthenia gravis. N/A=not available. QMG=Quantitative Myasthenia Gravis score.

treatment of relapsing remitting multiple sclerosis (ie, ocrelizumab, ofatumumab, and ublituximab), and ocrelizumab is also approved for primary progressive multiple sclerosis. The anti-CD19 antibody inebilizumab showed safety in a phase 1 clinical trial and is being investigated in later stage clinical trials.⁵⁸

Although B cell targeting, like several other therapies, has proven effective in preventing inflammation originating from the periphery in multiple sclerosis (considered to be a correlate of new MRI lesions and relapses), an increasing body of evidence suggests that disease progression can occur without relapses.⁵⁹ Several scenarios have been suggested to explain disease progression without apparent cellular infiltration from outside the blood-brain barrier. For example, derived from the observation that B cells can form meningeal follicles, tertiary lymphoid structures60 adjacent to the cortex, a plausible scenario entails continuous neuronal damage by locally secreted inflammatory cytokines, such as GM-CSF, IL-6, and lymphotoxin- α .³⁴ In addition to the clinical observation of ongoing disease progression without relapses, the notion that B cells are sustainably depleted in the periphery and the CSF, while oligoclonal bands persist in the CSF under therapy, supports the scenario that therapeutic antibodies are not able to sufficiently target tissue-resident B cells. Use of B celldepleting anti-CD19 CAR T cells that have demonstrated their ability to penetrate into tissue might be advantageous. The phase 2 clinical trial KYSA-7 testing anti-CD19 CAR T cells in 12 patients with progressive multiple sclerosis (NCT06138132) will shed more light on this issue.

Challenges in treating neuroimmunological disorders with CART cells

CAR T cells can cause severe side-effects in patients with cancer, particularly CRS, neurotoxic effects, and haematotoxicity. Established risk factors in this patient population include systemic inflammation (ie, CRP or increased ferritin concentrations), pre-existing neurological damage, older age, and residual tumour burden before infusion of CAR T cells.18,19,61 Although patients with autoimmune disorders typically have increased inflammatory activity and, in the case of neuroimmunological disorders, neurological pre-damage (figure 2A), good safety outcomes have been observed in admittedly small cohorts, without occurrence of higher grade CRS or neurotoxic effects.20 One reason for this safety profile could be the amount of target antigen, which in autoimmune disorders and neuroimmunological disorders is lower than in malignancies and might lead to reduced activation of CAR T cells and, thus, fewer side-effects. Additionally, the effects of long-term immunosuppression on the immune system and its responsiveness-for example, in the form of released IL-6—are still open questions that need to be addressed. As previously discussed, case numbers are small, and more trials are warranted to provide a more conclusive

Table 2: Clinical studies of the use of CART cells in patients with neuroimmunological disorders

picture; this is especially apparent in view of the heterogeneity in terms of underlying biology and clinical presentation of the autoimmune disorders and neuroimmunological disorders treated so far. To mitigate the potential toxic effects of CAR T cells, one strategy that is used is transient CAR expression through mRNA gene transfer, a method that has demonstrated promising outcomes in people with myasthenia gravis.41 Additionally, it will be necessary to reconsider risk factors and the according scores, such as the CAR-HEMATOTOX score¹⁹ for CAR-related haematotoxicity, that have been established in CAR T-cell therapy for malignancies. Monitoring strategies might also need to be adapted, particularly for neurotoxicity. Patients with conditions such as autoimmune encephalitis might be unable to provide responses necessary for assessing functional status using ICE or CARTOX scores. 62 Therefore, new clinical monitoring tools need to be developed for such patients, including continuous EEG-monitoring or routine (structural) brian MRI (figure 2C).63,64

A prerequisite for the efficacy of CAR T-cell therapy is their proximity to the target cells (figure 2C).⁶⁵ CAR T cells can be detected in the CSF of patients with cancer, regardless of neurotoxic effects.^{16,61} Initially, CNS involvement was an exclusion criterion for trials in lymphoma owing to concerns about neurotoxic effects.^{66,47} However, the efficacy of CAR T cells against CNS lymphomas is now well established.⁶⁷

Regarding the ideal target antigen, one size might not fit all. In fact, one potential explanation for the limited effectiveness of anti-CD20 antibody-mediated B-cell depletion is that autoantibody production might be attributed to CD20-negative plasmablasts or CD20negative plasma cells (either short-lived or long-lived variants). Plasmablasts express CD19, but plasma cells do not. Antigens shared by plasma cells and plasmablasts include CD38 and BCMA. Previous experience in systemic lupus erythematosus, myositis, or myasthenia gravis has shown that eliminating CD19positive plasmablasts, but not CD19-negative plasma cells, has proven sufficient to reach clinical response. 24,25,43 Reduction in disease activity was achieved without large depletion of IgG and increasing susceptibilty to infection in most patients. A more specific approach is the use of so-called chimeric autoantibody receptor (CAAR) T cells.68 Unlike CARs, which identify and attach to their target through an extracellular antibody fragment, CAARs direct the cytotoxic effects of T cells only to B cells producing autoantibodies, which has the advantage of reducing the risk of general immunosuppression. Because CAAR T cells also bind circulating autoantibodies, CAARs could become saturated, so a higher number of infused CAAR T cells than CAR T cells might be required to be effective. An additional potential risk of CAAR T-cell therapy is the so-called tip of the iceberg phenomenon in autoimmunity, which implies that all autoantibodies

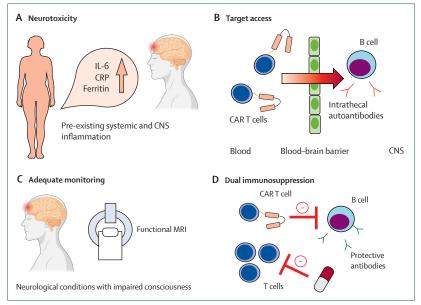


Figure 2: Potential challenges in the treatment of neuroimmunological disorders with CART cells

(A) Pre-existing inflammatory milieu and neurological disorders can be risk factors for therapy-associated toxic effects, such as cytokine release syndrome, neurotoxic effects, and haematotoxicity. (B) In some neuroimmunological disorders, autoantibodies are produced by B cells intrathecally. CART cells must overcome the blood-brain barrier to ensure effective treatment. (C) Because neuroimmunological disorders affect the nervous system, close neurological monitoring during and after CART-cell therapy is required to detect and manage potential neurological side-effects. New routines, such as functional MRI or continuous EEG monitoring, need to be developed, because reduced vigilance could render traditional clinical tests for assessing neurotoxicity obsolete. (D) Many neuroimmunological disorders require the use of immunosuppressants, affecting the T-cell compartment. Following B cell-targeting CART-cell therapy with consecutive depletion of B cells, such an approach should be carefully considered to avoid double immunosuppression with highly increased infection risk.

contributing to pathology are not always identified. To overcome this problem, at least partly, CAAR T cells with multiple specificities would be advantageous. However, it is possible that various neuroimmunological disorders will benefit differently from distinct approaches. Therefore, we must await the data from ongoing and future clinical trials (table 2).

The efficacy of CAR T cells naturally correlates with the fitness of the T cells (ie, their ability to expand, meet their metabolic demands, and kill their target cells). However, patients with neuroimmunological disorders are usually treated for long periods with immunosuppressive drugs, including those that target the T cell compartment, such as mycophenolate mofetil, which interferes with the function of the T cell, or natalizumab, which interferes with T cell-This immunosuppression should be trafficking. considered before lymphocyte apheresis and before implementing appropriate washout periods based on the drug's half-life and mechanism of action. Additionally, caution must be exercised with therapeutic T cell-targeted interventions following CAR T cell-mediated depletion of B cells. Simultaneous suppression of cellular and humoral immune responses could further increase the risk of infection (figure 2D). On the basis of the available data (which are limited), 69,70 it would be preferable to resume such therapy after the functional reconstitution of B cells if

disease activity permits or requires it. Another issue with autologous T cells is the risk that autoreactive T cells might be transfected in CAR T cell production. T cells directed against self-antigens such as MBP71 seem to have a role in neuroimmunological disorders, and expanding CAR T cells carrying an autoreactive endogenous TCR could theoretically worsen the clinical picture after infusion. One strategy to avoid both heavily pretreated, unfit T cells or autoreactive CAR T cells could be the use of so-called allogeneic T cells from third-party donors.11 To prevent graft-versus-host disease, the endogenous TCR is eliminated by genome editing or the immunogenicity is reduced by modifying MHC-I expression. It remains to be seen whether the initial success of allogeneic CAR T cells in malignant diseases can be replicated in autoimmune diseases.

Overall, therapies using genetically modified cell products are a multidisciplinary and complex effort involving at least neurologists and cell therapists (typically haematologists). Therefore, we refer readers to a position paper from the European Bone Marrow Transplantation's multidisciplinary working group on autoimmune diseases, which details the intended modus operandi of this collaboration between disciplines and the qualitative requirements.⁷²

Conclusions and future directions

The use of CAR T cells in the treatment of neuroimmunological disorders is expected to attract considerable attention in the coming years. Although data are scarce, the results so far are encouraging. This innovative approach could potentially result in extended periods of disease-free survival and treatment discontinuation. The pathophysiology of neuro-immunological disorders is complex and will require confirmation in pre-clinical models and controlled trials to determine if an immunological reset, similar to what has

Search strategy and selection criteria

We searched PubMed, medRxiv, bioRxiv, and Google Scholar for literature published between Jan 1, 2010, and March 1, 2024. Our search criteria included terms such as "CART cell" and "chimeric antigen receptor" and phrases targeting specific areas of interest, including "B-cell depletion", "B-cell targeting", "treatment and autoimmune disease", "treatment and autoimmune neurological disease", "treatment and neuroimmunological disease", and "treatment and neuroinflammation". We limited our search to papers published in English, and considered various types of publications, including clinical studies, mechanistic studies, case reports, case series, and review articles. In terms of cell therapy approaches, we have limited ourselves to CART cells and have not included studies of autologous or allogeneic stem cell transplantation or other types of adoptive cell therapies, such as regulatory T-cells or mesenchymal stromal cells.

been observed in autoimmune diseases, can be induced and under which specific conditions. When using CAR T cells in neuroimmunological disorders, safety should be carefully considered, particularly neurotoxicity. Patients' pre-existing neurological conditions, particularly those associated with CNS inflammation, might further increase the risk of CAR T cell-associated neurotoxicity and could also complicate monitoring for neurotoxicity. Several ongoing and planned studies are exploring the effects of CAR T cells in neuroimmunological disorders, offering an avenue for deeper insights into the potential of this innovative approach.

Contributors

AH and DM contributed equally to the writing, literature inclusion, and interpretation of the manuscript. GS helped writing the manuscript. AH had final responsibility to submit for publication.

Declaration of interests

AH has served on scientific advisory boards for Galapagos, Novartis, Merck Serono; has received speaker honoraria from Biogen Idec, Merck Serono, and Novartis; and has received research grants from Merck Serono. GS has received speaker honoraria from BMS, Cabaletta, Janssen, Kyverna, Miltenyi, and Novartis. DM has received speaker honoraria and consulting fees from Abbvie, BMS, Beigene, Celgene, Galapagos, Gilead, Janssen, Miltenyi, and Novartis.

Acknowledgments

AH is supported by the Deutsche Zentrum für Neurodegenerative Erkrankungen, the Deutsche Forschungsgemeinschaft (DFG) CRC Transregio 128 (Multiple Sklerose). GS is supported by the DFG through the Leibnitz Award and the research group PANDORA FOR2886, CRC1483 (EmpkinS), and CRC1181. GS is also supported by the Bundesministerium für Bildung und Forschung (Mascara project), the EU (ERC Synergy grant 4D Nanoscope), and the Innovative Medicines Initiatives (RTCure). DM is supported by the DFG through research group PANDORA FOR2886, CRC/TRR305 (A03), CRC/TRR221 (A06), and RTG2408 (P13) and grant 404074532.

References

- Cencioni MT, Mattoscio M, Magliozzi R, Bar-Or A, Muraro PA. B cells in multiple sclerosis—from targeted depletion to immune reconstitution therapies. *Nat Rev Neurol* 2021; 17: 399–414.
- 2 Cree BAC, Bennett JL, Kim HJ, et al. Inebilizumab for the treatment of neuromyelitis optica spectrum disorder (N-MOmentum): a double-blind, randomised placebo-controlled phase 2/3 trial. *Lancet* 2019; 394: 1352–63.
- 3 Maggi P, Bulcke CV, Pedrini E, et al. B cell depletion therapy does not resolve chronic active multiple sclerosis lesions. eBioMedicine 2023; 94: 104701.
- 4 June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* 2018; 359: 1361–65.
- 5 Jayaraman J, Mellody MP, Hou AJ, et al. CAR-T design: elements and their synergistic funtion. eBioMedicine 2020; 58: 102931.
- 6 Kanatas P, Stouras I, Stefanis L, Stathopoulos P. B-cell-directed therapies: a new era in multiple sclerosis treatment. Can J Neurol Sci 2023; 50: 355–64.
- 7 Kochenderfer JN, Rosenberg SA. Treating B-cell cancer with T cells expressing anti-CD19 chimeric antigen receptors. Nat Rev Clin Oncol 2013; 10: 267–76.
- 8 Abramson JS, Palomba ML, Gordon LI, et al. Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study. *Lancet* 2020; 396: 839–52.
- 9 Melenhorst JJ, Chen GM, Wang M, et al. Decade-long leukaemia remissions with persistence of CD4⁺ CAR T cells. *Nature* 2022; 602: 503–09.
- 10 Gumber D, Wang LD. Improving CAR-T immunotherapy: overcoming the challenges of T cell exhaustion. eBioMedicine 2022; 77: 103941.

- 11 Depil S, Duchateau P, Grupp SA, Mufti G, Poirot L. "Off-the-shelf" allogeneic CAR T cells: development and challenges. Nat Rev Drug Discov 2020; 19: 185–99.
- 12 Ghassemi S, Durgin JS, Nunez-Cruz S, et al. Rapid manufacturing of non-activated potent CAR T cells. *Nat Biomed Eng* 2022; 6: 118–28.
- 13 Amini L, Silbert SK, Maude SL, et al. Preparing for CAR T cell therapy: patient selection, bridging therapies and lymphodepletion. Nat Rev Clin Oncol 2022; 19: 342–55.
- 14 Brudno JN, Kochenderfer JN. Toxicities of chimeric antigen receptor T cells: recognition and management. *Blood* 2016; 127: 3321–30.
- Pensato U, Amore G, D'Angelo R, et al. Frontal predominant encephalopathy with early paligraphia as a distinctive signature of CAR T-cell therapy-related neurotoxicity. J Neurol 2022; 269: 609–15.
- 16 Gust J, Hay KA, Hanafi L-A, et al. Endothelial activation and bloodbrain barrier disruption in neurotoxicity after adoptive immunotherapy with CD19 CAR-T cells. *Cancer Discov* 2017; 7: 1404–19.
- Morris EC, Neelapu SS, Giavridis T, Sadelain M. Cytokine release syndrome and associated neurotoxicity in cancer immunotherapy. Nat Rev Immunol 2022; 22: 85–96.
- 18 Grant SJ, Grimshaw AA, Silberstein J, et al. Clinical presentation, risk factors, and outcomes of immune effector cell-associated neurotoxicity syndrome following chimeric antigen receptor T cell therapy: a systematic review. Transplant Cell Ther 2022; 28: 294–302.
- 19 Rejeski K, Perez A, Sesques P, et al. CAR-HEMATOTOX: a model for CAR T-cell-related hematologic toxicity in relapsed/refractory large B-cell lymphoma. *Blood* 2021; 138: 2499–513.
- Schett G, Mackensen A, Mougiakakos D. CAR T-cell therapy in autoimmune diseases. *Lancet* 2023; 402: 2034–44.
- 21 Weiner GJ. Rituximab: mechanism of action. Semin Hematol 2010; 47: 115–23.
- 22 Monson NL, Cravens PD, Frohman EM, Hawker K, Racke MK. Effect of rituximab on the peripheral blood and cerebrospinal fluid B cells in patients with primary progressive multiple sclerosis. Arch Neurol 2005; 62: 258–64.
- 23 Zamvil SS, Hauser SL. Antigen presentation by B cells in multiple sclerosis. N Engl J Med 2021; 384: 378–81.
- 24 Mougiakakos D, Kronke G, Volkl S, et al. CD19-targeted CAR T cells in refractory systemic lupus erythematosus. N Engl J Med 2021; 385: 567–69.
- 25 Muller F, Boeltz S, Knitza J, et al. CD19-targeted CAR T cells in refractory antisynthetase syndrome. *Lancet* 2023; 401: 815–18.
- 26 Bergmann C, Müller F, Distler JHW, et al. Treatment of a patient with severe systemic sclerosis (SSc) using CD19-targeted CAR T cells. Ann Rheum Dis 2023; 82: 1117–20.
- 27 Muller F, Taubmann J, Bucci L, et al. CD19 CAR T-cell therapy in autoimmune disease—a case series with follow-up. N Engl J Med 2024; 390: 687–700.
- 28 Mullard A. CAR T cell therapies raise hopes—and questions—for lupus and autoimmune disease. *Nat Rev Drug Discov* 2023; 22: 859–61.
- 29 Halliley JL, Tipton CM, Liesveld J, et al. Long-lived plasma cells are contained within the CD19⁻CD38ʰCD138⁺ subset in human bone marrow. *Immunity* 2015; 43: 132–45.
- 30 Ingwersen J, Masanneck L, Pawlitzki M, et al. Real-world evidence of ocrelizumab-treated relapsing multiple sclerosis cohort shows changes in progression independent of relapse activity mirroring phase 3 trials. Sci Rep 2023; 13: 15003.
- 31 Saccà F, Barnett C, Vu T, et al. Efgartigimod improved healthrelated quality of life in generalized myasthenia gravis: results from a randomized, double-blind, placebo-controlled, phase 3 study (ADAPT). J Neurol 2023; 270: 2096–105.
- 32 Lee DSW, Rojas OL, Gommerman JL. B cell depletion therapies in autoimmune disease: advances and mechanistic insights. Nat Rev Drug Discov 2021; 20: 179–99.
- 33 Barr TA, Shen P, Brown S, et al. B cell depletion therapy ameliorates autoimmune disease through ablation of IL-6producing B cells. J Exp Med 2012; 209: 1001–10.
- 34 Li R, Rezk A, Miyazaki Y, et al. Proinflammatory GM-CSFproducing B cells in multiple sclerosis and B cell depletion therapy. Sci Transl Med 2015; 7: 310ra166.

- 35 Ransohoff RM. Multiple sclerosis: role of meningeal lymphoid aggregates in progression independent of relapse activity. *Trends Immunol* 2023; 44: 266–75.
- 36 Verschuuren JJ, Palace J, Murai H, Tannemaat MR, Kaminski HJ, Bril V. Advances and ongoing research in the treatment of autoimmune neuromuscular junction disorders. *Lancet Neurol* 2022; 21: 189–202.
- 37 Marino M, Bartoccioni E, Alboini PE, Evoli A. Rituximab in myasthenia gravis: a "to be or not to be" inhibitor of T cell function. Ann N Y Acad Sci 2018; 1413: 41–48.
- 38 Howard JF, Utsugisawa K, Benatar M, et al. Safety and efficacy of eculizumab in anti-acetylcholine receptor antibody-positive refractory generalised myasthenia gravis (REGAIN): a phase 3, randomised, double-blind, placebo-controlled, multicentre study. *Lancet Neurol* 2017; 16: 976–86.
- 39 Vu T, Meisel A, Mantegazza R, et al. Terminal complement inhibitor ravulizumab in generalized myasthenia gravis. NEJM Evid 2022; published online April 26. https://doi.org/10.1056/ EVIDoa2100066.
- 40 Howard JF, Bril V, Vu T, et al. Safety, efficacy, and tolerability of efgartigimod in patients with generalised myasthenia gravis (ADAPT): a multicentre, randomised, placebo-controlled, phase 3 trial. *Lancet Neurol* 2021; 20: 526–36.
- 41 Granit V, Benatar M, Kurtoglu M, et al. Safety and clinical activity of autologous RNA chimeric antigen receptor T-cell therapy in myasthenia gravis (MG-001): a prospective, multicentre, openlabel, non-randomised phase 1b/2a study. *Lancet Neurol* 2023; 22: 578–90.
- 42 Chahin N, Sahagian G, Feinberg MH, et al. Twelve-month followup of patients with generalized myasthenia gravis receiving BCMAdirected mRNA cell therapy. *medRxiv* 2024; published online Jan 4. https://doi.org/10.1101/2024.01.03.24300770 (preprint).
- 43 Haghikia A, Hegelmaier T, Wolleschak D, et al. Anti-CD19 CAR T cells for refractory myasthenia gravis. *Lancet Neurol* 2023; 22: 1104–05.
- 44 Tian DS, Qin C, Dong MH, et al. B cell lineage reconstitution underlies CAR-T cell therapeutic efficacy in patients with refractory myasthenia gravis. EMBO Mol Med 2024; 16: 966–87.
- 45 Qin C, Dong M-H, Zhou L-Q, et al. Single-cell analysis of refractory anti-SRP necrotizing myopathy treated with anti-BCMA CAR-T cell therapy. Proc Natl Acad Sci USA 2024; 121: e2315990121.
- 46 Pittock SJ, Berthele A, Fujihara K, et al. Eculizumab in aquaporin-4-positive neuromyelitis optica spectrum disorder. N Engl J Med 2019; 381: 614–25.
- 47 Schuster SJ, Bishop MR, Tam CS, et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. N Engl J Med 2019; 380: 45–56.
- 48 Jarius S, Aktas O, Ayzenberg I, et al. Update on the diagnosis and treatment of neuromyelits optica spectrum disorders (NMOSD) revised recommendations of the Neuromyelitis Optica Study Group (NEMOS). Part I: Diagnosis and differential diagnosis. *J Neurol* 2023; 270: 3341–68.
- 49 Banwell B, Bennett JL, Marignier R, et al. Diagnosis of myelin oligodendrocyte glycoprotein antibody-associated disease: International MOGAD Panel proposed criteria. *Lancet Neurol* 2023; 22: 268–82.
- 50 Barreras P, Vasileiou ES, Filippatou AG, et al. Long-term effectiveness and safety of rituximab in neuromyelitis optica spectrum disorder and MOG antibody disease. *Neurology* 2022; 99: e2504–16.
- 51 Nosadini M, Eyre M, Molteni E, et al. Use and Safety of immunotherapeutic management of N-methyl-D-aspartate receptor antibody encephalitis: a meta-analysis. *JAMA Neurol* 2021; 78: 1333-44.
- 52 Smets I, Titulaer MJ. Antibody therapies in autoimmune encephalitis. *Neurotherapeutics* 2022; **19**: 823–31.
- 53 Pittock SJ, Barnett M, Bennett JL, et al. Ravulizumab in aquaporin-4-positive neuromyelitis optica spectrum disorder. *Ann Neurol* 2023; 93: 1053–68.
- 54 Traboulsee A, Greenberg BM, Bennett JL, et al. Safety and efficacy of satralizumab monotherapy in neuromyelitis optica spectrum disorder: a randomised, double-blind, multicentre, placebocontrolled phase 3 trial. *Lancet Neurol* 2020; 19: 402–12.

- 55 Qin C, Tian D-S, Zhou L-Q, et al. Anti-BCMA CAR T-cell therapy CT103A in relapsed or refractory AQP4-IgG seropositive neuromyelitis optica spectrum disorders: phase 1 trial interim results. Signal Transduct Target Ther 2023; 8: 5.
- 56 Bar-Or A, Li R. Cellular immunology of relapsing multiple sclerosis: interactions, checks, and balances. *Lancet Neurol* 2021; 20: 470–83.
- 57 Jain RW, Yong VW. B cells in central nervous system disease: diversity, locations and pathophysiology. *Nat Rev Immunol* 2022; 22: 513–24.
- 58 Agius MA, Klodowska-Duda G, Maciejowski M, et al. Safety and tolerability of inebilizumab (MEDI-551), an anti-CD19 monoclonal antibody, in patients with relapsing forms of multiple sclerosis: results from a phase 1 randomised, placebo-controlled, escalating intravenous and subcutaneous dose study. Mult Scler 2019; 25: 235–45.
- 59 Tur C, Carbonell-Mirabent P, Cobo-Calvo Á, et al. Association of early progression independent of relapse activity with long-term disability after a first demyelinating event in multiple sclerosis. *JAMA Neurol* 2023; 80: 151–60.
- 60 Magliozzi R, Howell OW, Calabrese M, Reynolds R. Meningeal inflammation as a driver of cortical grey matter pathology and clinical progression in multiple sclerosis. *Nat Rev Neurol* 2023; 19: 461–76.
- 61 Santomasso BD, Park JH, Salloum D, et al. Clinical and biological correlates of neurotoxicity associated with CAR T-cell therapy in patients with B-cell acute lymphoblastic leukemia. *Cancer Discov* 2018; 8: 958–71.
- 62 Pennisi M, Jain T, Santomasso BD, et al. Comparing CAR T-cell toxicity grading systems: application of the ASTCT grading system and implications for management. *Blood Adv* 2020; 4: 676–86.
- 63 Herlopian A, Dietrich J, Abramson JS, Cole AJ, Westover MB. EEG findings in CAR T-cell therapy-related encephalopathy. Neurology 2018; 91: 227–29.
- 64 Stoecklein S, Wunderlich S, Papazov B, et al. Functional connectivity MRI provides an imaging correlate for chimeric antigen receptor T-cell-associated neurotoxicity. *Neurooncol Adv* 2023; published online Oct 24. https://doi.org/10.1093/noajnl/vdad135.

- 65 Pruss H. Autoantibodies in neurological disease. Nat Rev Immunol 2021: 21: 798–813.
- 66 Neelapu SS, Locke FL, Bartlett NL, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. N Engl J Med 2017; 377: 2531–44.
- 67 Cook MR, Dorris CS, Makambi KH, et al. Toxicity and efficacy of CAR T-cell therapy in primary and secondary CNS lymphoma: a meta-analysis of 128 patients. *Blood Adv* 2023; 7: 32–39.
- 68 Reincke SM, von Wardenburg N, Homeyer MA, et al. Chimeric autoantibody receptor T cells deplete NMDA receptor-specific B cells. Cell 2023; 186: 5084–97.
- 69 Mankikian J, Caille A, Reynaud-Gaubert M, et al. Rituximab and mycophenolate mofetil combination in patients with interstitial lung disease (EVER-ILD): a double-blind, randomised, placebocontrolled trial. Eur Resp J 2023; 61: 2202071.
- 70 Kridin K, Mruwat N, Amber KT, Ludwig RJ. Risk of infections in patients with pemphigus treated with rituximab vs. azathioprine or mycophenolate mofetil: a large-scale global cohort study, Br J Dermatol 2023; 188: 499–505.
- 71 Burns J, Rosenzweig A, Zweiman B, Lisak RP. Isolation of myelin basic protein-reactive T-cell lines from normal human blood. Cell Immunol 1983; 81: 435–40.
- 72 Greco R, Alexander T, Del Papa N, et al. Innovative cellular therapies for autoimmune diseases: expert-based position statement and clinical practice recommendations from the EBMT practice harmonization and guidelines committee. eClinicalMedicine 2024; 69: 102476

Copyright © 2024 The Author(s). Published by Elsevier Ltd. This is an Open Access article under the CC BY 4.0 license.