

REVIEW

Open Access



Longitudinal biomarker studies in human neuroimaging: capturing biological change of Alzheimer's pathology

Larissa Fischer^{1,2*} , Dana Parker^{2†} , Samira Maboudian³ , Corrina Fonseca³ , Claudia Tato-Fernández⁴ , Lucie Annen^{5,6}, Prithvi Arunachalam^{7,8} , Julia R. Bacci⁹ , Michelle Barbour¹⁰ , Serena Capelli¹¹ , Stamatia Karagianni^{12,13} , Lyduine E. Collij^{7,14} , Paul Edison^{15,16}, Nick C. Fox^{17,18} , Nicolai Franzmeier^{13,19,20} , Michel J. Grothe^{12,21} , William J. Jagust³ , Anne Maass^{1,22} , Maura Malpetti^{23,24} , Ross W. Paterson^{17,18} , Aitana Sogorb-Esteve^{17,18}  and Michael Schöll^{12,13,17*} 

Abstract

Despite extensive research, open questions about the biological underpinnings of Alzheimer's disease (AD) remain. Neuroimaging biomarkers based on positron emission tomography (PET) and magnetic resonance imaging (MRI) offer in vivo insights into these complex biological changes and interactions. However, most evidence to date comes from cross-sectional studies, limiting our understanding of disease progression. Longitudinal studies enable the investigation of biological changes within individuals, revealing how pathology evolves over time. With this review, we provide an overview of how longitudinal imaging biomarker studies have advanced the field and how they can contribute to future research. We highlight longitudinal biomarker studies that have provided critical insights into disease trajectories, staging, and individual variability. We further assess longitudinal multimodal studies which have elucidated interactions between AD-specific pathology, amyloid- β and tau, and broader biological changes like neurodegeneration, neuronal dysfunction, vascular disease, and inflammation. Further, we discuss associations of brain changes with symptomatology and clinical outcomes and conclude with challenges and future directions.

Keywords Alzheimer's disease, Biomarker, Longitudinal, Neuroimaging, PET, MRI

[†]Larissa Fischer and Dana Parker contributed equally to this work.

*Correspondence:

Larissa Fischer
larissa.fischer@dzne.de
Michael Schöll
michael.scholl@neuro.gu.se

Full list of author information is available at the end of the article



Introduction

Alzheimer's disease (AD) is characterized by a cascade of biological changes, particularly the accumulation of amyloid- β (A β) and tau pathology, which progressively affect neuronal functioning and integrity. Pathology impairs cognitive abilities and eventually leads to AD dementia. Insights into the pathophysiology of AD have been gained from a variety of biomarkers, with human postmortem histopathological studies as the gold standard. A biomarker is "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention" [1]. In recent decades, neuroimaging biomarkers have been developed using positron emission tomography (PET) to assess A β and tau burden, as well as PET and magnetic resonance imaging (MRI) to evaluate nonspecific pathophysiological changes such as neurodegeneration, network dysfunction, vascular disease, and inflammation [2, 3].

Human neuroimaging has the important benefit of providing region-specific insights into changes of the underlying biology in an in vivo setting, but the field has been largely dominated by cross-sectional studies. Consequently, most of the evidence for the proposed model of AD pathophysiological sequences has relied on cross-sectional imaging or histology studies. However, as the field has grown, information from longitudinal and multimodal imaging studies is becoming increasingly available. With repeated imaging over time, those studies can provide a more detailed understanding of the temporospatial development of AD, uncover mechanisms contributing to individual variability in disease trajectories, and can further help to narrow down clinically relevant biomarkers. These studies not only allow inferences about the dynamics of disease progression, but also help to identify potential underlying mechanisms affecting the disease process and outcomes. When incorporating interventions, these studies are further uniquely poised to identify causal relationships and can provide insights into how the brain reacts to novel therapies.

The purpose of this review is to assess the utility of longitudinal neuroimaging studies in capturing the biological changes along the AD continuum and their interactions with each other. First, we will assess what aspects of the underlying pathology are captured by human neuroimaging. We will then examine what longitudinal imaging biomarkers can tell us about (1) the disease trajectory and pathological staging in AD, (2) relationships between AD-specific pathology and other nonspecific biological changes observed in AD (neurodegeneration, neuronal dysfunction, vascular disease, and inflammation), and (3) how these interrelated biological changes are linked to symptomatology and clinical outcomes. Finally, we conclude by discussing potential imaging biomarkers

that currently lack longitudinal support, highlighting challenges and possible insights from future studies. By focusing on the advances and challenges in longitudinal imaging biomarkers of AD, this review ultimately aims to provide insights into the biological underpinnings of AD that could contribute to improved tools for diagnosis and disease monitoring, as well as determining suitable treatment targets to attenuate AD progression.

Disease trajectory and pathological staging

First, we provide a brief overview of AD pathology and imaging biomarkers before discussing longitudinal trajectories and staging. AD is associated with diverse pathological changes that can be captured with neuroimaging methods in humans, each reflecting distinct yet interacting biological processes. The core AD features, discovered in postmortem histological research, are extracellular A β plaques and intraneuronal tau neurofibrillary tangles [4–7]. Both can be measured in vivo using PET imaging, which captures the buildup of these protein aggregates in the brain [8, 9]. Further, nonspecific pathological changes related to AD encompass neurodegeneration and neuronal dysfunction, including changes in brain metabolism and networks, and changes related to vascular disease and inflammation. An overview is provided in Table 1.

Amyloid pathology

Neuritic A β plaques have long been recognized as a histopathological hallmark of AD, with early diffuse neocortical plaques depositing in the posteromedial cortex (PMC) and frontal regions. Characteristic hierarchical stages ("Thal phases") were established by postmortem histology [6]. This staging has been largely recapitulated with A β -PET imaging [25], which has enabled investigating early emerging amyloidosis in cognitively normal individuals and accumulation over time. There is a spatiotemporal hierarchy of A β accumulation [8, 26, 27] and longitudinal A β progression patterns closely match cross-sectional staging [28–30]. Rates of A β deposition show very little variability across anatomically distant brain regions [31] and resemble sigmoid-shaped trajectories, with higher global A β burden at baseline predicting higher rates of neocortical A β accumulation in both cognitively unimpaired and impaired individuals and with accumulation slowing down at higher levels of A β accumulation [32, 33]. While soluble A β oligomers may spread across neighboring regions, plaque formation could rather depend on local factors like intense neuronal activity [34, 35].

Tau pathology

Tau tangles are closely related to cognition [36], first deposit in the (trans)entorhinal cortex, and accumulate

Table 1 Imaging of biological changes related to Alzheimer's disease

Pathological change	How can we measure this change with imaging?	What underlying biological feature or process is targeted?
Amyloid- β (A β) accumulation	Positron emission tomography (PET) imaging using, for example, the tracers [11C]Pittsburgh compound B (PiB), [18F]florbetaben (FBB), [18F]florbetapir (FBP), and [18F]flutemetamol [8–10]	Insoluble A β plaque accumulation surrounding neurons is a specific feature of Alzheimer's disease (AD) and can be assessed by specific tracers binding to these proteins [6, 7]
Tau accumulation	PET imaging using, for example, the tracers [18F]flortaucipir (FTP), [18F]PM-PBB3/florzolotau, [18F]MK-6240, and [18F]PI-2620 [11, 12]	Tau neurofibrillary tangle accumulation within neurons is a specific feature of AD and can be assessed by specific tracers binding to tau aggregates [4, 5]
Neurodegeneration	T1- and T2-weighted structural MRI (sMRI) focusing on the gray matter volume of brain structures, FLAIR sequences, and diffusion-weighted imaging (DWI) focusing on white matter microstructural connectivity	Brain atrophy, white matter hyperintensities (WMH), and white matter impairment are disruptions commonly found over the course of AD. Hippocampal atrophy assessed with sMRI is a key prognostic feature of AD [13]. Widespread abnormalities in white matter microstructure have been consistently reported in DWI studies of patients with AD [14]
Neuronal dysfunction	Fluorodeoxyglucose (FDG) and synaptic vesicle protein 2A (SV2A) PET using the [11C]UCB-J or [18F]SynVest-1 tracer, fMRI studies using the blood oxygenation level dependent (BOLD) method, perfusion PET, SPECT, and MRI using e.g. the arterial spin labeling (ASL) MRI sequence	Changes in glucose brain metabolism measured via FDG-PET is an indicator of neuronal activity [15]. SV2A-PET imaging tracers binding to the SV2A protein aims at investigating synaptic integrity [16]. Functional imaging using BOLD fMRI is an indirect measure of network dysfunction using the magnetic properties of oxygenated blood [17]. Perfusion MRI using labeling of arterial blood water as an endogenous tracer for blood flow and perfusion PET and SPECT using radiotracers [18]
Vascular disease	T1-weighted sMRI and FLAIR sequences Perfusion MRI to investigate cerebral perfusion abnormalities using dynamic contrast enhanced (DCE) MRI	WMH and enlarged perivascular spaces (PVS) are biomarkers for small vessel disease (SVD) and used to investigate the separate and joint influence of SVD and AD pathology on the disease course [19]. WMH might be of vascular or non-vascular origin [20]. Blood–brain-barrier (BBB) integrity might be reflected in cerebral perfusion abnormalities [21]
Inflammation	PET tracers 18 kDa translocator protein (TSPO) and Deuterium-L-deprenyl (DED)	TSPO-PET signal most likely reflects microglia density [22, 23], [11C]DED-PET aims to visualize activated astrocytes [24]

A β Amyloid-beta, AD Alzheimer's Disease, ASL Arterial Spin Labeling, BOLD Blood Oxygenation Level Dependent, DCE Dynamic Contrast Enhanced, DED Deuterium-L-deprenyl, DWI Diffusion Weighted Imaging, FBB [18F]florbetaben, FBP [18F]florbetapir, FDG Fluorodeoxyglucose, fMRI functional Magnetic Resonance Imaging, FTP [18F]flortaucipir, PET Positron Emission Tomography, PiB Pittsburgh Compound B, PVS Perivascular Spaces, sMRI Structural Magnetic Resonance Imaging, SPECT Single Photon Emission Computed Tomography, SV2A Synaptic Vesicle Protein 2A, SVD=Small Vessel Disease, TSPO 18 kDa Translocator Protein, WMH White Matter Hyperintensities

throughout the medial temporal lobe (MTL). In the presence of elevated A β , tau subsequently progresses to temporoparietal regions and finally across the neocortex. This pattern was first characterized in postmortem tissue samples [4] but has been confirmed in vivo in cross-sectional PET studies [37–39]. A priori region-based studies also suggest that tau generally accumulates in these patterns longitudinally [40, 41] but show considerable individual variability in tau deposition and spread [39, 42–45]. Tau spread along structural [46] and functional [42] connections has also been observed longitudinally. Further, higher rates of tau deposition in the MTL are predicted by locally higher baseline tau burden in cognitively unimpaired older adults and may further be driven by local activity [47]. While rates of tau accumulation were similar across brain regions in one study [48], another study reported higher rates of accumulation for temporal regions [49] in cognitively unimpaired and impaired adults. Additionally, data-driven profiling has identified fast accumulators with increased accumulation in temporal cortex and PMC [45].

Neurodegeneration and neuronal dysfunction

Structural MRI (sMRI) has played an integral role in investigating AD progression and diagnosis. Whole-brain and hippocampal atrophy are sensitive markers of neurodegeneration and disease progression [50, 51]. At the whole-brain level, a classical "cortical signature" of AD-related atrophy is well-established and associated with symptom severity [52]. Longitudinal sMRI studies have further demonstrated that rates of gray matter loss in AD compared to elderly controls generally mirror patterns of tau accumulation [53–55] and precede symptomatic onset in both familial [56, 57] and sporadic AD [58]. However, despite these associations, atrophy is not specific for AD pathology; for example, hippocampal atrophy is associated with cognitive decline independent of A β and tau pathology, suggesting contributions from other pathological factors [59]. Further, studies showing increased gray matter volume or cortical thickness with early A β , possibly related to glia response [60], and "pseudatrophy" in anti-amyloid trials [61, 62] call sMRI into question as being a universal marker of neurodegeneration.

FDG-PET is a widely used imaging modality for assessing region-specific aberrant brain glucose metabolism

related to AD pathology. Its prognostic utility lies in its ability to detect early region-specific hypometabolism that correlates with cognitive decline before clinical symptoms become apparent [63–65]. While FDG-PET and sMRI are often used interchangeably as imaging biomarkers in AD [2], evidence from multimodal studies suggests that FDG-PET is more sensitive to early neurodegenerative processes compared to sMRI [66–68]. Moreover, the extent and pattern of hypometabolism correlate with advancing AD pathology, providing a means to track disease severity over time [69, 70] and conduct clinical classification [71, 72]. Interestingly, there are differential FDG-PET patterns related to different underlying pathologies, which can provide clinically useful information for differential diagnosis [73, 74].

Modalities that target neuronal integrity, namely SV2A-PET and diffusion weighted imaging (DWI), seem to be more closely associated with tau than A β pathology. Longitudinal SV2A-PET studies remain scarce, but synaptic loss over time has been shown to follow tau rather than A β accumulation patterns [75, 76] and diffusion tensor imaging (DTI) studies have been linked to axonal integrity and show that it is particularly impacted by tau pathology, preceding both neuronal loss and clinical manifestation [77–80]. More recent advances in DWI, such as multi-shell acquisitions, allow a more detailed investigation of region-specific subtle microstructural dysfunction, providing the potential for early detection of AD [81]. Overall, however, regional onsets and spatio-temporal progression of AD-specific patterns using these modalities are still incompletely understood.

Longitudinal BOLD fMRI studies in AD typically focus on resting-state functional connectivity (FC), while longitudinal studies on task-based FC and activity are rare [17]. Using fMRI, early functional changes like “hyperactivation” and “hyperconnectivity” linked to AD pathology and cognition have been identified and are interpreted as markers of dysfunctional brain networks [82]. fMRI studies can bridge molecular and clinical research by shedding light on network mechanisms of risk and resilience to AD pathology [83–85]. However, most fMRI studies use a group approach rather than precision imaging as they were designed to contribute to cognitive neuroscience research rather than to explain between-subject variance [86, 87]. Moreover, BOLD signal changes are not specific to AD and occur in normal aging and various neurodegenerative diseases [88–90].

To summarize, MRI and FDG-PET approaches add valuable information to understand altered brain responses related to AD pathology and its progression and relationship with cognitive symptoms. However, while FDG-PET is an established marker of neurodegeneration and can be used to stage disease progression, it does not directly measure A β or tau pathology and

therefore cannot alone determine neuropathological stage. Similarly, structural and functional MRI provide important but indirect measures of underlying pathology. Combining these modalities with molecular imaging or other biomarkers offers a more complete and biologically specific picture of disease progression.

Pathological interactions and potential causality **Longitudinal characterization of the pathological cascade of Alzheimer's disease**

The classic model of AD biomarker change from normal aging along the AD continuum influenced research over the last decade greatly. It suggests that A β and tau accumulate up to 20 years before clinical manifestation [32, 91]. In this model, A β accumulation is seen as a very early, potentially initiating factor in the cascade of AD [7, 92], enabling tau spread, which in turn leads to synaptic and neuronal loss [93]. The cascading network failure model of AD [94, 95] further incorporates higher local activity of the default mode network (DMN) and higher between-network connectivity. It is debated whether these functional changes initially serve as compensatory processes for decreasing network function related to early AD pathology. However, they could also reflect oversaturation of brain networks which, in turn, leads to accelerated network failure. These complex theoretical models are largely based on cross-sectional data, and it is difficult to empirically address causality. Extensive longitudinal multimodal studies with participants from healthy adults to severe stages of AD including interventions would be critical to address the issue. Longitudinal multimodal studies have, however, contributed insights into parts of the temporal dynamics of AD.

An established finding is that A β drives tau accumulation and spread. Tau accumulation rates are elevated with higher A β burden in diverse brain areas [48]. Recent longitudinal studies showed that A β facilitates tau spread from medial to lateral temporal lobe and neocortical regions [47, 96, 97]. Conversely, higher baseline tau in temporal and parietal cortex was associated with faster A β accumulation [31].

Regarding neurodegeneration, higher superior-temporal but not global A β burden predicted greater cortical thinning in patients with mild cognitive impairment (MCI) but not in cognitively unimpaired adults [98]. In another study of cognitively unimpaired adults, however, higher A β burden at baseline predicted a steeper decline in hippocampal volume [99] and in white matter integrity of the parahippocampal cingulum, while there was no association between baseline measures [100]. Critically, longitudinal studies suggest that tau drives neurodegeneration more strongly than A β . Baseline global tau- but not A β -PET signal predicted the rate and topography of prospective atrophy in dementia patients [55].

In cognitively unimpaired older adults, the steepest rate of tau accumulation and atrophy has been reported in temporal and retrosplenial cortex, in dementia patients, however, regions differed, with the steepest rate of tau accumulation in frontal cortex and atrophy in PMC [101]. Frontotemporal cortical thinning has been found to be predicted by higher baseline tau burden, but not by change in tau-PET signal, in cognitively unimpaired and impaired individuals [102].

Baseline tau pathology also predicts faster synaptic loss as measured by SV2A-PET [75], and synaptic loss regionally follows tau-accumulation patterns over time [76], indicating that tau is implicated in synaptic loss. Tau pathology may also drive unfavorable functional changes. A recent study using longitudinal fMRI during encoding and cerebrospinal fluid (CSF)-markers of AD pathology proposed that MTL atrophy and tau accumulation are independently linked to reduced deactivations in the DMN, which includes the PMC [103]. Further, tau might mediate the association of A β and neurodegeneration [104], and conversely, A β might mediate the association of tau and neurodegeneration. Studies report that abnormal hippocampal cingulum bundle diffusivity at baseline predicts tau accumulation in the PMC only in A β -positive individuals [99]. In A β -positive individuals, increase in cortical tau has been further found to be related to a diffuse increase in atrophy in frontotemporo-parietal areas, while increase in A β itself is not [49]. While the complex causal relationships along the AD cascade are still not fully understood, multimodal studies combining longitudinal biomarkers can advance our

understanding of temporal dynamics beyond the current simplified models (see Fig. 1B).

Role of network dysfunction

Network dysfunction may play a central role regarding the spatiotemporal dynamics of AD pathology. MTL and PMC hyperactivation could predispose those brain regions to pathology accumulation (i.e. tau in MTL and A β in PMC) and contribute to accelerated spread of pathology [17] (see Fig. 2). Further, transneuronal tau spread from the MTL to neocortical regions might be accelerated via aberrant functional connectivity [105, 106]. However, these models are largely based on animal or human cross-sectional studies, and the interplay with microstructural changes is unclear [107]. Recent longitudinal multimodal studies have begun to reveal how network changes in AD relate to pathology accumulation and spread.

Longitudinal studies in cognitively unimpaired older adults using memory task-fMRI suggest that higher and increasing BOLD signal, especially of the hippocampus, predicts the accumulation of A β and tau. More specifically, higher hippocampal but not frontal or occipital fMRI activation during successful encoding predicts increased accumulation of global A β [108] and local fMRI activity predicts increased accumulation of MTL tau [47]. Regarding the PMC, increasing precuneus activation over time during episodic retrieval relates to higher subsequent global A β -PET burden in *APOE4* carriers [109]. Further, increase [110] as well as decrease [111] in DMN resting-state FC (rsFC) has been related to faster A β accumulation, indicating failure of the DMN

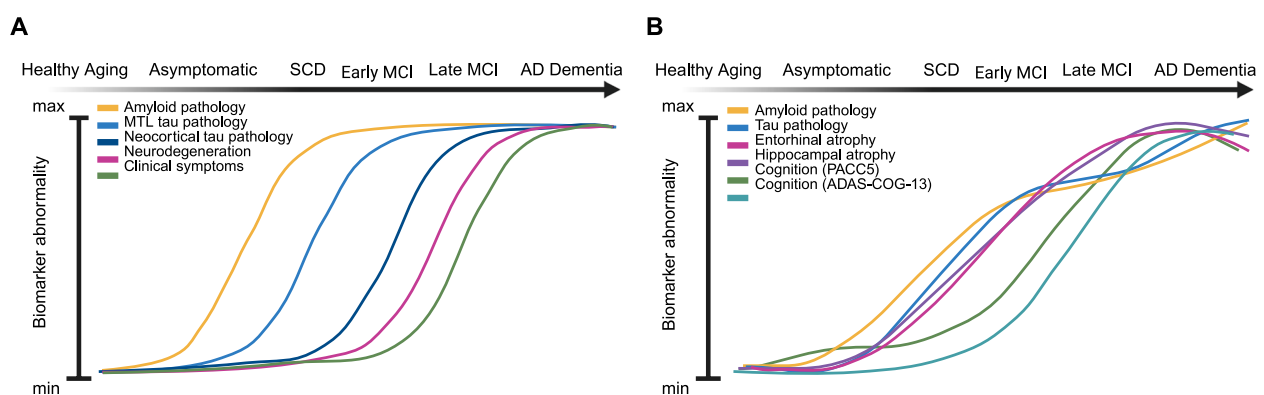


Fig. 1 Conceptual illustration of longitudinal biomarker dynamics in Alzheimer's disease. **A** The influential model of Jack and colleagues [2] depicts archetypical sigmoidal curves representing isolated changes in Alzheimer's disease biomarkers over time, based on the revised AT(N) framework. Adapted from [2]. **B** We propose that moving from isolated biomarker studies to longitudinal multimodal investigations can uncover more complex interactions and causal relationships between biomarkers. The curves shown in B are adapted from a longitudinal modeling study by Lattmann-Greve and colleagues [103], illustrating how multimodal longitudinal data can reveal intricate and interacting dynamics over time. In their study, the authors utilized longitudinal CSF, MRI, and cognitive scores in a multivariate probabilistic disease progression model to generate empirical biomarker disease progression curves. The resulting curves uncovered differential hypothetically implicated biomarker trajectories with cognition being preceded by morphometry and CSF-based Alzheimer's disease biomarkers, respectively, and different timepoints of fastest change. The authors further assessed the relationship to change in fMRI encoding task activation. These changes in activation were nonlinear and independently associated with tau positivity and neurodegeneration. Adapted from [103]

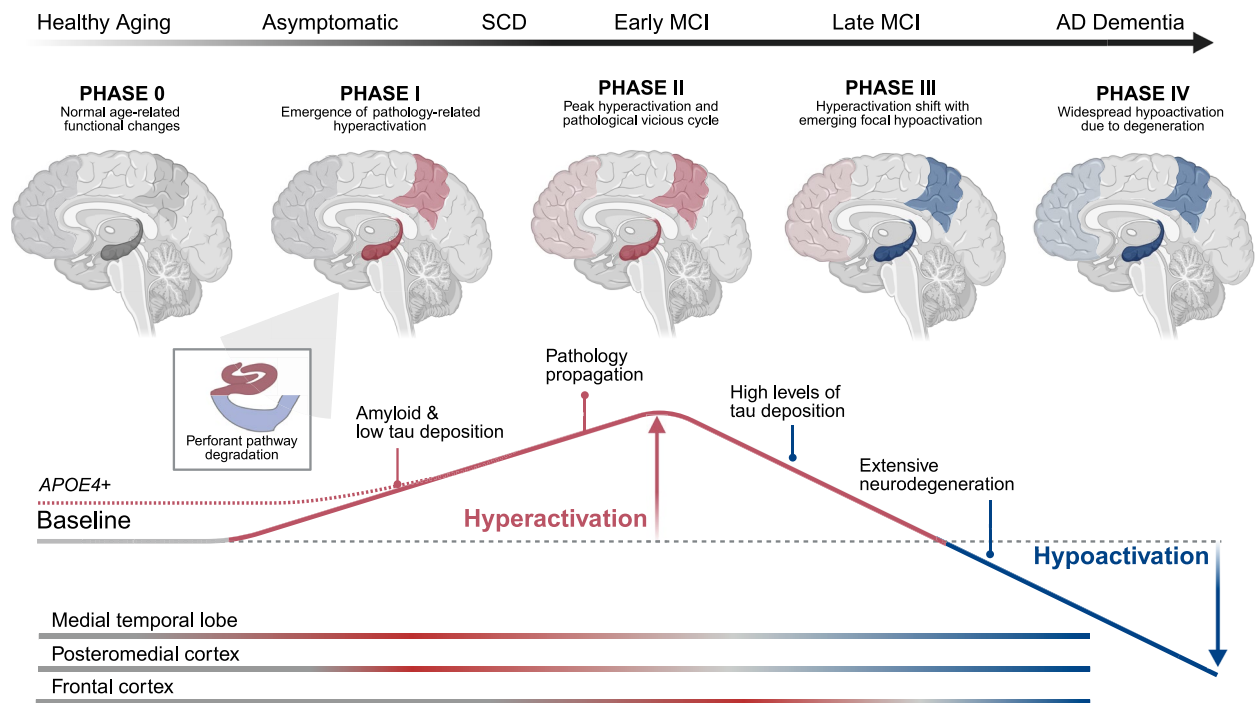


Fig. 2 Proposed model of hyper- and hypoactivation in the Alzheimer's disease pathological cascade. In Phase 0, non-pathological aging is characterized by functional changes (baseline, grey) in comparison with younger adults. Genetic predisposition to Alzheimer's disease (AD) (i.e. *APOE4* genotype) may cause a prolonged state of increased activation across mid- to late life (red dotted line). In Phase I, age- and/or genetic-related functional changes predispose certain regions to pathology accumulation (i.e. hyperphosphorylated tau in medial temporal lobe (MTL) and A β in posteromedial cortex (PMC)). This pathology accumulation coincides with the emergence of task-based hyperactivation (red), defined as increased activation contrasted against healthy older adults, which is evident when probed with episodic memory tasks. Hyperactivation first occurs in the hippocampus, particularly within dentate gyrus/CA3, due to tau-related perforant path degeneration (see inset box) and in PMC regions due to A β -related effects. Overt memory impairment is not yet evident at this stage. In Phase II, disconnection between the MTL and PMC results in exaggerated hyperactivation, as well as accelerated expansion of pathology in a vicious cycle. This peak of hyperactivation is associated with SCD and early MCI. In Phase III, a tipping point of high levels of tau pathology ultimately leads to neuronal silencing and neurodegeneration, resulting in hypoactivation (blue) which first emerges in the hippocampus and PMC. Simultaneously, a shift in hyperactivation to other regions (e.g. frontal cortex) occurs. Finally, in Phase IV, widespread pathology and neurodegeneration leads to further hypoactivation that encompasses large-scale cortical regions and networks, resulting in overt cognitive impairment characteristic of AD dementia. Adapted from [17]

system as a critical precursor of spatiotemporal A β progression. Moreover, aberrant FC could drive tau spread. Baseline hippocampal tau predicts precuneus tau accumulation, particularly when higher rsFC between those regions and higher baseline A β burden are present [112]. Pathology-related higher bidirectional effective connectivity of the DMN and MTL during repetition of stimuli predicts entorhinal tau accumulation [113] and increasing within-hippocampus rsFC has been associated with plasma p-tau increase in *APOE4* carriers [114]. More specifically, tau seems to spread along functional connections. Findings from animal models show that tau spreads transneuronally from the MTL to neocortical regions [115]. Longitudinal human fMRI studies suggest the same process in humans, with aberrantly higher FC patterns accelerating tau spread [40, 105, 106, 116].

Role of vascular disease

Although vascular dysregulation has long been acknowledged as an important contributor to AD pathology [117,

118], it is often overlooked in prevailing AD models [32]. However, longitudinal studies suggest that vascular dysregulation may be among the earliest pathological events in AD, highlighting its importance for early intervention and therapeutic development [119, 120].

Often considered a surrogate marker of small vessel disease (SVD), white matter hyperintensity (WMH) volumes have been linked to vascular dysfunction and dysregulation early in the process of AD [121]. However, emerging evidence highlights the heterogeneity of WMH pathophysiology, suggesting that WMH might also be caused by AD-related neurodegeneration and inflammation [20]. Longitudinal studies showed that WMH volume increase is associated with increase in A β -PET signal, hippocampal atrophy, and cortical thinning in elderly controls [122] and that WMH burden predicts increased hippocampal atrophy in elderly controls and MCI patients [123]. WMH progression and cortical atrophy may be mutually reinforcing processes, as individuals with higher baseline WMH volumes experience

faster cortical thinning in temporal, cingulate and insular regions, and individuals with lower initial cortical thickness experience more rapid WMH progression in these regions [124]. The interplay between A β and WMH is complex. While A β deposition can exacerbate WMH burden through mechanisms like neuroinflammation and oxidative stress [125], WMH themselves may accelerate A β pathology by impairing clearance mechanisms [126], creating a vicious cycle amplifying pathology. A longitudinal study over eight years showed that higher WMH burden is associated with an increase in A β accumulation in cognitively unimpaired individuals [127]. This bidirectional relationship underscores the potential for WMH to mediate the impact of A β on clinical outcome, independent of traditional vascular risk factors such as hypertension.

A further biomarker for vascular contributions to AD is perivascular space (PVS) enlargement. Longitudinal studies are still rare, but recently, higher burden of cerebral microvascular lesions predicted faster progression of PVS enlargement [128]. While CSF A β -positivity is linked to PVS volume increase in the centrum semiovale, combined A β - and tau-positivity is associated with basal ganglia PVS volume increase [129].

Role of inflammation

A β plaques are surrounded by activated microglia, indicating a strong relationship between the pathological progression of AD and inflammation [130–133]. Microglia migrate to A β lesions and are related to the degradation of A β peptides and the clearance of A β [133]. The role of microglia in causing or responding to AD pathology is still being debated [134] due to microglial cells having both protective as well as neurotoxic phenotypes [135].

To date the only confirmed visualization method of activated microglia and inflammation is PET, with cross-sectional studies using the 18kD translocator protein (TSPO) tracer [23] dominating the field. Alternative tracers are under development, such as [11C]DED-PET to assess reactive astrogliosis, which demonstrates higher binding at early stages of AD [24]. Neuroinflammation increases in AD, demonstrated by higher TSPO levels throughout the cortex, particularly in fronto-temporal regions [136]. Microglial activation is related to tau pathology and cognitive decline in symptomatic patients [137–139] but might be more closely related to A β burden in the absence of cognitive symptoms [140], an effect that could be modulated by the *APOE4* genotype [141]. Thus, an early peak in cortical TSPO binding might be a response to A β deposition, whereas a second peak in temporal regions could reflect tau propagation.

Longitudinal TSPO-PET studies have shown that neuroinflammation increases over time in AD [130, 142],

correlating with cognitive impairment [142]. Increasing microglial activation over time appears to be directly related to A β and inversely related to glucose metabolism in AD [130]. However, neuroinflammation is a dynamic process and there might be different profiles of microglial activation that cannot be differentiated with TSPO-PET and may have a distinct impact on disease progression.

Relationships between biomarker changes and cognition

Longitudinal A β -PET imaging studies have demonstrated that faster A β accumulation is modestly correlated with global cognitive decline over short follow-up times [143] and is linked to progression from being cognitively unimpaired to MCI over eight to ten years [110]. Recent studies also suggest that longitudinal A β accumulation is more closely related to changes in non-memory domains rather than episodic memory, particularly in A β -positive cognitively unimpaired individuals and MCI patients [143–146]. This association could be related to the tendency of A β to accumulate multifocally across the cortex and affect functional circuits responsible for coordinating multiple cognitive functions. Furthermore, these studies suggest that the rate of A β accumulation is more influential on cognitive changes at earlier clinical stages along the AD continuum. Additionally, the spatial extent of A β could be a more sensitive measure for cognition than A β levels [147]. In contrast, longitudinal tau-PET studies show that MTL and early neocortical tau accumulation are more strongly associated with episodic memory change [144] and clinical outcomes [36] than A β . Though this relationship is significant in adults with low A β burden, the association is enhanced in A β -positive individuals and significant regardless of concurrent atrophy. This suggests that early tau accumulation, especially when influenced by elevated A β , may affect cognition through mechanisms other than atrophy, such as inflammation, microstructural or metabolic changes [148–150]. Longitudinal sMRI and DWI studies have, however, shown that atrophy and microstructural changes are linked to cognition and clinical outcomes in AD [151–153]. Increases particularly in hippocampal atrophy are associated with faster decline in episodic memory in cognitively unimpaired individuals [154] and in symptomatic AD [56]. Clinical impairment is related to widespread decreases in fractional anisotropy and increases in mean diffusivity, reflecting microstructural white matter degeneration [14].

Further, metabolic and functional changes are related to cognitive decline. Longitudinal decreases in metabolism measured using FDG-PET are linked to global cognitive decline and predict cognitive instability [69, 155, 156] and decreases in ASL-measured whole-brain perfusion are related to decline in processing speed in cognitively

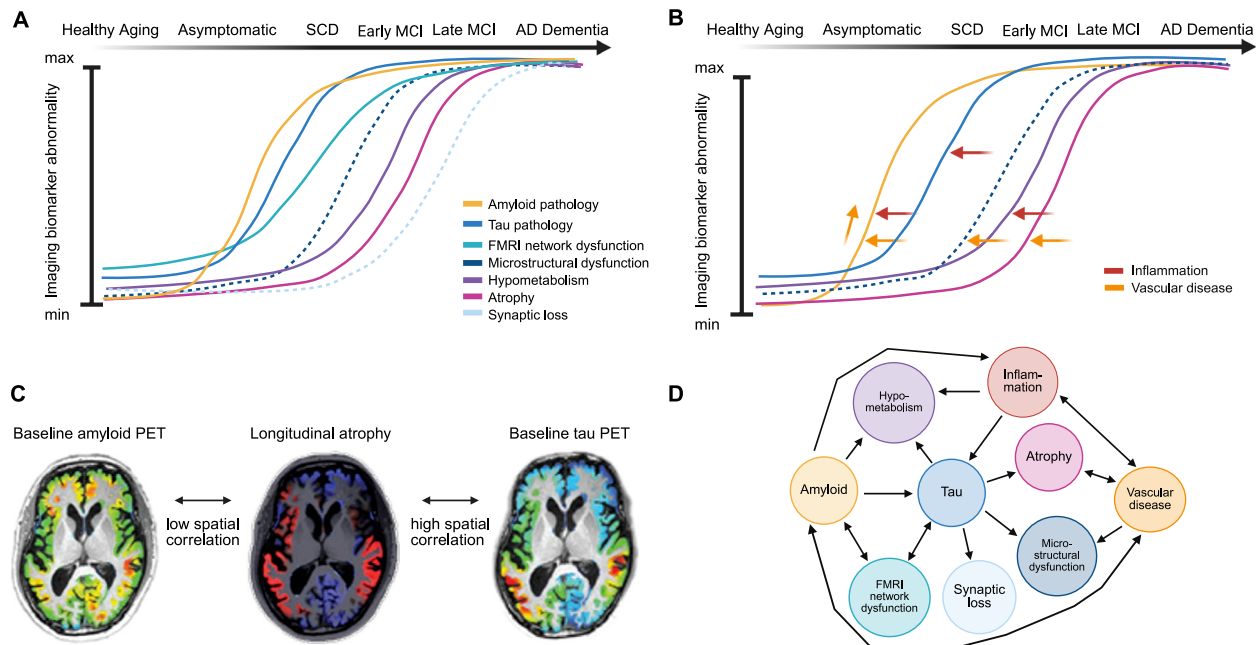


Fig. 3 Conceptual illustration of the insights gained from longitudinal multimodal imaging biomarker studies on temporal, spatial, and causal aspects of Alzheimer's disease pathology. This figure is not meant to be exhaustive but serves to illustrate the complex interplay of Alzheimer's disease imaging biomarkers over time. **A** Temporal trajectories and relationships of imaging biomarkers across the disease continuum, derived from longitudinal studies discussed in this review paper. Curves depict the estimated onset, rate of change, and plateau phases for biomarkers. Dotted lines indicate biomarkers where limited longitudinal data is available. **B** Arrows depict shifts in biomarker trajectories influenced by inflammation and vascular disease, emphasizing how these additional factors alter disease trajectory. **C** Spatial correspondence of pathological processes across brain regions, illustrating patterns of colocalization and divergence as assessed by La Joie and colleagues [55] using multimodal longitudinal imaging biomarkers, can provide valuable insight into disease dynamics. Adapted from [55]. Brain plots from [55]. Reprinted with permission from AAAS. **D** Graph of causal relationships between imaging biomarkers based on studies reviewed above. Nodes represent distinct pathological processes implicated in Alzheimer's disease. Directed edges indicate putative causal influences between processes, as estimated from longitudinal observational and experimental data to date as discussed in this review

unimpaired individuals [157]. Longitudinal fMRI studies have played a crucial role in identifying functional changes, such as specific regional activation and network connectivity patterns, that are related to early cognitive changes in AD. For example, in cognitively unimpaired individuals and A β -positive MCI patients, higher hippocampal activity during encoding predicts decline in global cognition [158, 159]. Similarly, the absence of hyperactivation in the precuneus during a recognition task is associated with better episodic memory performance in *APOE4* non-carriers [109]. Resting-state studies suggest early increases in connectivity between the MTL and cortical regions and the default mode network with AD pathology, which is also associated with decline in global cognition and episodic memory [88, 160, 161].

Notably, there is a more pronounced cognitive decline with vascular co-pathology. Longitudinal increase in WMH volume is steeper over the age of 60 and associated with a more rapid cognitive decline [121, 122]. Highlighting the dynamic nature of WMH, progression of WMH is related to decline, while regression and stability of WMH is related to improvement in cognition [162].

Taken together, tau accumulation is closely related to domain-specific memory decline, as well as functional

changes involving the MTL-PMC episodic memory network measured with fMRI. A β -PET, FDG-PET, sMRI, and DWI provide valuable biomarkers to predict global cognition and clinical outcomes. However, many longitudinal cohort studies that focus on biomarkers only have a limited range of cognitive tests in their assessment, often only a coarse measure of global cognition (e.g. MMSE, MoCA) and it remains open which biomarkers can capture (future) change in more fine-grained cognitive functions.

Biomarkers lacking current longitudinal investigation: challenges and potential future insights

While there is robust longitudinal data for A β and tau pathology that has been contributing to a better understanding of the mechanisms behind AD, other biological features, such as neuroinflammation, vascular changes, and synaptic integrity, remain underexplored, despite recent efforts and advances (see Fig. 3 for a schematic overview). These processes may present significant factors in disease progression, but it is not yet fully understood how they evolve over time. Although longitudinal studies remain the gold standard for establishing the

temporal sequence of disease-related changes, emerging data-driven approaches such as SuStaIn (Subtype and Stage Inference)[163] can help infer likely progression patterns from cross-sectional datasets, providing valuable insights when longitudinal data are lacking (see [164] for a review).

A major challenge is the lack of suitable PET tracers. Sufficiently specific PET tracers for alpha-synuclein co-pathology [165, 166] are lacking, tracers such as [18F]flortaucipir bind well to 3R/4R but do not bind equally well to other tauopathies [167], and SV2A-PET assessment needs to be further validated. 11C-UCB-J is an effective PET tracer for SV2A and provides insights into synaptic density, however, it is important to recognize that it is an indirect measure of synaptic density [168, 169]. The tracer binds specifically to SV2A, a protein found in pre-synaptic vesicles, but this binding reflects the presence of synaptic vesicles rather than a direct count of synapses themselves. Longitudinal studies combining SV2A-PET with FDG-PET, fMRI and sMRI measures could generate joint topographical maps of change, contributing to a better understanding of the underlying biological processes. Advancements in tracer development can thus open up exciting new avenues for multimodal imaging research.

A second major challenge is the limited understanding of factors that accelerate AD progression and mechanisms underlying resilience and resistance. A key question regarding disease acceleration is whether vascular pathology represents an independent process or whether it is pathophysiologically connected to A β and tau [129, 170, 171]. Longitudinal alterations in WMH and PVS need further exploration to understand how their rate of change relates to core AD markers and cognition. Future longitudinal studies should therefore investigate the regional relationship between rate of change in WMH, A β and tau deposition to elucidate interactions. Further, some older adults harboring AD-pathology can stay cognitively unimpaired for longer than expected given the severity of pathology [172, 173]. Longitudinal imaging and cognitive assessment combined with post-mortem histology can shed light on mechanisms of resilience and resistance across scales [174].

A third major challenge is the lack of longitudinal data from diverse cohorts [175]. Cohorts that better reflect societal heterogeneity are crucial to better understand the complex role of socio-economic, ethno-racial and demographic factors that influence the trajectory of AD [176–178]. Further, they can pave the way to better address interindividual differences in modifiable risk factors for AD [179]. Collecting longitudinal data of diverse cohorts could therefore be a valuable aim in clinical trials for novel treatments [180]. Vice-versa, investigating these rich longitudinal datasets from clinical intervention

studies can offer opportunities to infer causal relationships of disease mechanisms.

Thus, developing and validating imaging biomarkers, disentangling the contribution of co-pathologies to the trajectory of AD, and using rich datasets are central goals for future studies to better understand disease mechanisms and foster clinical advancement [181, 182].

Conclusion

To conclude, the unique insights into AD gained from longitudinal imaging studies highlight their importance as a key direction for future research. Longitudinal human neuroimaging biomarker studies are suited to capture the temporospatial dynamics of biological changes along the Alzheimer's continuum. By tracking changes over time, they can offer a deeper understanding of complex interacting processes like disease acceleration by co-pathology. Particularly longitudinal multimodal imaging can reveal joint evolving patterns of e.g. tau accumulation, synaptic loss and metabolic changes that cross-sectional studies cannot detect, helping to refine our understanding of disease progression and offering more accurate predictions of symptom development. Particularly when focusing on refined PET tracers and diverse cohorts, the gained insights allow for a more comprehensive perspective on the development and interplay of different pathologies, which is crucial for both early diagnosis and the evaluation of therapeutic interventions.

Abbreviations

A β	Amyloid-beta
AD	Alzheimer's Disease
ASL	Arterial Spin Labeling
BOLD	Blood Oxygenation Level Dependent
DCE	Dynamic Contrast Enhanced
DED	Deuterium-L-deprenyl
DMN	Default Mode Network
DWI	Diffusion Weighted Imaging
DTI	Diffusion Tensor Imaging
FBB	[18F]florbetaben
FBP	[18F]florbetapir
FC	Functional Connectivity
FDG	Fluorodeoxyglucose
fMRI	Functional Magnetic Resonance Imaging
FTP	[18F]flortaucipir
MTL	Medial Temporal Lobe
PET	Positron Emission Tomography
PIB	Pittsburgh Compound B
PMC	Posteromedial Cortex
PVS	Perivascular Spaces
sMRI	Structural Magnetic Resonance Imaging
SV2A	Synaptic Vesicle Protein 2A
SVD	Small Vessel Disease
TSPO	18KDa Translocator Protein
WMH	White Matter Hyperintensities

Acknowledgements

We thank the faculty of the University of Gothenburg/UCL/BBRC course "Biomarkers in Neurodegenerative Diseases" for sharing their expertise and for the stimulating discussions that inspired this review.

Authors' contributions

Conceptualization: LF, DP, SM, CF, MS. Figures: LF. Literature review: LF, DP, SM, CF, CT-F, LA, PA, JRB, MB, SC, SK. Guidance and supervision: LEC, NF, MJG, WJJ, AM, MM, RWP, AS-E, MS. Writing original draft: LF, DP, SM, CF, CT-F. Writing – review and editing: All authors. All authors read and approved the final manuscript.

Funding

Open access funding provided by University of Gothenburg. This research did not receive any specific grant from funding agencies in the public, commercial or not-for-profit sectors. Open access funding was provided by the University of Gothenburg.

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

- ¹German Center for Neurodegenerative Diseases (DZNE), Magdeburg, Germany
- ²Department of Neurobiology and Behavior, University of California, Irvine, USA
- ³Department of Neuroscience, University of California, Berkeley, USA
- ⁴Turku PET Centre, University of Turku, Turku University Hospital, Turku, Finland
- ⁵Division of Geriatric Psychiatry, University Hospitals of Geneva, Thônex, Switzerland
- ⁶Department of Psychiatry, University of Geneva, Geneva, Switzerland
- ⁷Department of Radiology and Nuclear Medicine, UMC Vrije Universiteit Amsterdam, Amsterdam, The Netherlands
- ⁸Amsterdam Neuroscience, Brain Imaging, Amsterdam, The Netherlands
- ⁹Department of Epidemiology and Prevention, Wake Forest University School of Medicine, Winston-Salem, USA
- ¹⁰Alzheimer Center, Department of Neurology, UMC Vrije Universiteit Amsterdam, Amsterdam, the Netherlands
- ¹¹Bioengineering Department, Istituto di Ricerche Farmacologiche Mario Negri IRCCS, Ranica, Italy
- ¹²Wallenberg Centre for Molecular and Translational Medicine, University of Gothenburg, Sahlgrenska University Hospital, Gothenburg, Sweden
- ¹³Department of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, The Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden
- ¹⁴Clinical Memory Research Unit, Department of Clinical Sciences, Faculty of Medicine, Lund University, Malmö, Sweden
- ¹⁵Division of Neurology, Department of Brain Sciences, Faculty of Medicine, Imperial College London, London, UK
- ¹⁶School of Medicine, Cardiff University, Cardiff, UK
- ¹⁷Dementia Research Centre, UCL Queen Square Institute of Neurology, University College London, London, UK
- ¹⁸UK Dementia Research Institute at UCL, University College London, London, UK
- ¹⁹Institute for Stroke and Dementia Research (ISD), University Hospital, LMU Munich, Munich, Germany
- ²⁰Munich Cluster for Systems Neurology (SyNergy), Munich, Germany
- ²¹Reina Sofia Alzheimer Centre, CIEN Foundation, ISCIII, Madrid, Spain
- ²²Faculty of Natural Sciences, Otto Von Guericke University Magdeburg, Magdeburg, Germany
- ²³Department of Clinical Neurosciences and Cambridge University Hospitals NHS Trust, University of Cambridge, Cambridge, UK
- ²⁴UK Dementia Research Institute at University of Cambridge, Cambridge, UK

Received: 27 May 2025 / Accepted: 21 November 2025

Published online: 04 December 2025

References

1. Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Ther.* 2001;69:89–95.
2. Jack CR Jr, Andrews JS, Beach TG, Buracchio T, Dunn B, Graf A, et al. Revised criteria for diagnosis and staging of Alzheimer's disease: Alzheimer's Association workgroup. *Alzheimer's & Dementia.* 2024;20:5143–69.
3. Márquez F, Yassa MA. Neuroimaging biomarkers for Alzheimer's disease. *Mol Neurodegener.* 2019;14:21.
4. Braak H, Braak E. Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol.* 1991;82:239–59.
5. Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol.* 2006;112:389–404.
6. Thal DR, Rüb U, Orantes M, Braak H. Phases of a beta-deposition in the human brain and its relevance for the development of AD. *Neurology.* 2002;58:1791–800.
7. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med.* 2016;8:595–608.
8. Grothe MJ, Barthel H, Sepulcre J, Dyrba M, Sabri Q, Teipel SJ, et al. In vivo staging of regional amyloid deposition. *Neurology.* 2017;89:2031–8.
9. Chételat G, La Joie R, Villain N, Perrotin A, de La Sayette V, Eustache F, et al. Amyloid imaging in cognitively normal individuals, at-risk populations and preclinical Alzheimer's disease. *NeuroImage: Clinical.* 2013;2:356–65.
10. Heurling K, Leuzy A, Zimmer ER, Lubberink M, Nordberg A. Imaging β -amyloid using [(18)F]flutemetamol positron emission tomography: from dosimetry to clinical diagnosis. *Eur J Nucl Med Mol Imaging.* 2016;43:362–73.
11. Maass A, Berron D, Harrison TM, Adams JN, La Joie R, Baker S, et al. Alzheimer's pathology targets distinct memory networks in the ageing brain. *Brain.* 2019;142:2492–509.
12. Gérard T, Colmant L, Malotau V, Salman Y, Huyghe L, Quenon L, et al. The spatial extent of tauopathy on [F]MK-6240 tau PET shows stronger association with cognitive performances than the standard uptake value ratio in Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2024;51:1662–74.
13. Dutton RA, Dinov ID, Hayashi KM, Toga AW, Cummings JL, Thompson PM. Conversion of mild cognitive impairment to Alzheimer disease predicted by hippocampal atrophy maps. *Arch Neurol.* 2006;63:693–9.
14. Mayo CD, Mazerolle EL, Ritchie L, Fisk JD, Gawryluk JR. Longitudinal changes in microstructural white matter metrics in Alzheimer's disease. *NeuroImage Clinical.* 2016;13:330–8.
15. Chételat G, Arbizu J, Barthel H, Garibotto V, Law I, Morbelli S, et al. Amyloid-PET and 18F-FDG-PET in the diagnostic investigation of Alzheimer's disease and other dementias. *Lancet Neurol.* 2020;19(11):951–62.
16. Kumar A, Scarpa M, Nordberg A. Tracing synaptic loss in Alzheimer's brain with SV2A PET-tracer UCB-J. *Alzheimer's & Dementia.* 2024;20:2589–605.
17. Corriveau-Lecavalier N, Adams JN, Fischer L, Molloy EN, Maass A. Cerebral hyperactivation across the Alzheimer's disease pathological cascade. *Brain Commun.* 2024;6:fcae376.
18. Lindner T, Bolar DS, Achten E, Barkhof F, Bastos-Leite AJ, Detre JA, et al. Current state and guidance on arterial spin labeling perfusion MRI in clinical neuroimaging. *Magn Reson Med.* 2023;89:2024–47.
19. Kalaria RN, Akinyemi R, Ihara M. Does vascular pathology contribute to Alzheimer changes? *J Neurol Sci.* 2012;322:141–7.
20. Garnier-Crussard A, Cotton F, Krolak-Salmon P, Chételat G. White matter hyperintensities in Alzheimer's disease: beyond vascular contribution. *Alzheimer's & Dementia.* 2023;19:3738–48.
21. Kerkhofs D, Wong SM, Zhang E, Staals J, Jansen JFA, van Oostenbrugge RJ, et al. Baseline blood-brain barrier leakage and longitudinal microstructural tissue damage in the periphery of white matter hyperintensities. *Neurology.* 2021;96:e2192.
22. Nutma E, Fancy N, Weinert M, Tsartsalis S, Marzin MC, Muirhead RCJ, et al. Translocator protein is a marker of activated microglia in rodent models but not human neurodegenerative diseases. *Nat Commun.* 2023;14:5247.
23. Wijesinghe SS, Rowe JB, Mason HD, Allinson KSJ, Thomas R, Vontobel DS, et al. Post-mortem validation of in vivo TSPO PET as a microglial biomarker. *Brain.* 2025;148:1904–10. <https://doi.org/10.1093/brain/awaf078>.

24. Chiotis K, Johansson C, Rodriguez-Vieitez E, Ashton NJ, Blennow K, Zetterberg H, et al. Tracking reactive astrogliosis in autosomal dominant and sporadic Alzheimer's disease with multi-modal PET and plasma GFAP. *Mol Neurodegener.* 2023;18:1–14.
25. Thal DR, Beach TG, Zhanette M, Lijia J, Heurling K, Chakrabarty A, et al. Estimation of amyloid distribution by [18F]flutemetamol PET predicts the neuropathological phase of amyloid β -protein deposition. *Acta Neuropathol.* 2018;136:557–67.
26. Fantoni E, Collij L, Alves IL, Buckley C, Farrar G. The spatial-temporal ordering of amyloid pathology and opportunities for PET imaging. *J Nucl Med.* 2020;61:166–71.
27. Mattsson N, Palmqvist S, Stomrud E, Vogel J, Hansson O. Staging β -amyloid pathology with amyloid positron emission tomography. *JAMA Neurol.* 2019;76:1319–29.
28. Jelicstratova I, Teipel SJ, Grothe MJ. Longitudinal validity of PET-based staging of regional amyloid deposition. *Hum Brain Mapp.* 2020;41:4219–31.
29. Collij LE, Heeman F, Salvadó G, Ingala S, Altomare D, de Wilde A, et al. Multitracer model for staging cortical amyloid deposition using PET imaging. *Neurology.* 2020;95:e1538–53.
30. Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K. Association between longitudinal plasma neurofilament light and neurodegeneration in patients with Alzheimer disease. *JAMA Neurol.* 2019;76:791–9.
31. LaPoint MR, Baker SL, Landau SM, Harrison TM, Jagust WJ. Rates of β -amyloid deposition indicate widespread simultaneous accumulation throughout the brain. *Neurobiol Aging.* 2022;115:1–11.
32. Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, et al. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol.* 2013;12(2):207–16.
33. Villain N, Chételat G, Grassiot B, Bourgeat P, Jones G, Ellis KA, et al. Regional dynamics of amyloid- β deposition in healthy elderly, mild cognitive impairment and Alzheimer's disease: a voxelwise PIB-PET longitudinal study. *Brain.* 2012;135:2126–39.
34. Pignataro A, Middei S. Trans-synaptic spread of amyloid- β in Alzheimer's disease: paths to β -amyloidosis. *Neural Plast.* 2017;2017:5281829.
35. Chen GF, Xu TH, Yan Y, Zhou YR, Jiang Y, Melcher K, et al. Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol Sin.* 2017;38:1205–35.
36. Hanseeuw BJ, Betensky RA, Jacobs HIL, Schultz AP, Sepulcre J, Becker JA, et al. Association of amyloid and tau with cognition in preclinical Alzheimer disease: a longitudinal study. *JAMA Neurol.* 2019;76:915–24.
37. Soleimani-Meigooni DN, Iaccarino L, La Joie R, Baker S, Bourakova V, Boxer AL, et al. 18F-flortaucipir PET to autopsy comparisons in Alzheimer's disease and other neurodegenerative diseases. *Brain.* 2020;143:3477–94.
38. Schöll M, Lockhart SN, Schonhaut DR, O'Neil JP, Janabi M, Ossenkoppele R, et al. Pet imaging of tau deposition in the aging human brain. *Neuron.* 2016;89:971–82.
39. St-Onge F, Chappelle M, Breitner JCS, Villeneuve S, Pichet Binette A. Tau accumulation and its spatial progression across the Alzheimer's disease spectrum. *Brain Commun.* 2024;6:fcae031.
40. Roemer-Cassiano SN, Wagner F, Evangelista L, Rauchmann BS, Dehsarvi A, Steward A, et al. Amyloid-associated hyperconnectivity drives tau spread across connected brain regions in Alzheimer's disease. *Sci Transl Med.* 2025;17:eadp2564.
41. Frontzkowski L, Ewers M, Brendel M, Biel D, Ossenkoppele R, Hager P, et al. Earlier Alzheimer's disease onset is associated with tau pathology in brain hub regions and facilitated tau spreading. *Nat Commun.* 2022;13:4899.
42. Franzmeier N, Dewenter A, Frontzkowski L, Dichgans M, Rubinski A, Neitzel J, et al. Patient-centered connectivity-based prediction of tau pathology spread in Alzheimer's disease. *Sci Adv.* 2020;6:eabd1327.
43. Vogel JW, Young AL, Oxtoby NP, Smith R, Ossenkoppele R, Strandberg OT, et al. Four distinct trajectories of tau deposition identified in Alzheimer's disease. *Nat Med.* 2021;27:871–81.
44. Leuzy A, Binette AP, Vogel JW, Klein G, Borroni E, Tonietto M, et al. Comparison of group-level and individualized brain regions for measuring change in longitudinal tau positron emission tomography in Alzheimer disease. *JAMA Neurol.* 2023;80:614–23.
45. Tosun D, Thropp P, Southehal S, Spottiswoode B, Fahmi R. Profiling and predicting distinct tau progression patterns: an unsupervised data-driven approach to flortaucipir positron emission tomography. *Alzheimer's & Dementia.* 2023;19:5605–19.
46. Yang F, Chowdhury SR, Jacobs HIL, Sepulcre J, Wedeen VJ, Johnson KA, et al. Longitudinal predictive modeling of tau progression along the structural connectome. *Neuroimage.* 2021;237:118126.
47. Adams JN, Harrison TM, Maass A, Baker SL, Jagust WJ. Distinct factors drive the spatiotemporal progression of tau pathology in older adults. *J Neurosci.* 2022;42:1352–61.
48. Jack CR Jr, Wiste HJ, Schwarz CG, Lowe VJ, Senjem ML, Vemuri P, et al. Longitudinal tau PET in ageing and Alzheimer's disease. *Brain.* 2018;141(5):1517–28.
49. Cho H, Choi JY, Lee HS, Lee JH, Ryu YH, Lee MS, et al. Progressive tau accumulation in Alzheimer disease: 2-year follow-up study. *J Nucl Med.* 2019;60:1611–21.
50. Frisoni GB, Fox NC, Jack CR Jr, Scheltens P, Thompson PM. The clinical use of structural MRI in Alzheimer disease. *Nat Rev Neurol.* 2010;6:67–77.
51. Fox NC, Warrington EK, Freeborough PA, Hartikainen P, Kennedy AM, Stevens JM, et al. Presymptomatic hippocampal atrophy in Alzheimer's disease. A longitudinal MRI study. *Brain.* 1996;119(Pt 6):2001–7.
52. Dickerson BC, Bakkour A, Salat DH, Feczko E, Pacheco J, Greve DN, et al. The cortical signature of Alzheimer's disease: regionally specific cortical thinning relates to symptom severity in very mild to mild AD dementia and is detectable in asymptomatic amyloid-positive individuals. *Cereb Cortex.* 2009;19:497–510.
53. Thompson PM, Hayashi KM, de Zubicaray G, Janke AL, Rose SE, Semple J, et al. Dynamics of Gray Matter Loss in Alzheimer's Disease. *J Neurosci.* 2003;23:994–1005.
54. Fjell AM, Walhovd KB, Fennema-Notestine C, McEvoy LK, Hagler DJ, Holland D, et al. One-year brain atrophy evident in healthy aging. *J Neurosci.* 2009;29:15223–31.
55. La Joie R, Visani AV, Baker SL, Brown JA, Bourakova V, Cha J, et al. Prospective longitudinal atrophy in Alzheimer's disease correlates with the intensity and topography of baseline tau-PET. *Sci Transl Med.* 2020;12:eaa5732.
56. Schott JM, Fox NC, Frost C, Scahill RI, Janssen JC, Chan D, et al. Assessing the onset of structural change in familial Alzheimer's disease. *Ann Neurol.* 2003;53:181–8.
57. Knight WD, Kim LG, Douiri A, Frost C, Rossor MN, Fox NC. Acceleration of cortical thinning in familial Alzheimer's disease. *Neurobiol Aging.* 2011;32:1765–73.
58. Sluimer JD, van der Flier WM, Karas GB, van Schijndel R, Barnes J, Boyes RG, et al. Accelerating regional atrophy rates in the progression from normal aging to Alzheimer's disease. *Eur Radiol.* 2009;19:2826–33.
59. Hanseeuw BJ, Jacobs HIL, Schultz AP, Buckley RF, Farrell ME, Guehl NJ, et al. Association of pathologic and volumetric biomarker changes with cognitive decline in clinically normal adults. *Neurology.* 2023;101:e2533–44.
60. Salvadó G, Shekari M, Falcon C, Operto G, Milà-Alomà M, Sánchez-Benavides G, et al. Brain alterations in the early Alzheimer's continuum with amyloid- β , tau, glial and neurodegeneration CSF markers. *Brain Commun.* 2022;4:fca134.
61. Belder CRS, Boche D, Nicoll JAR, Jaunmuktane Z, Zetterberg H, Schott JM, et al. Brain volume change following anti-amyloid β immunotherapy for Alzheimer's disease: amyloid-removal-related pseudo-atrophy. *Lancet Neurol.* 2024;23:1025–34.
62. Barkhof F, Knopman DS. Brain shrinkage in anti- β -amyloid Alzheimer trials: neurodegeneration or pseudoatrophy? *Neurology.* 2023;100:941–2.
63. Iaccarino L, Sala A, Perani D, Alzheimer's Disease Neuroimaging Initiative. Predicting long-term clinical stability in amyloid-positive subjects by FDG-PET. *Ann Clin Transl Neurol.* 2019;6:1113–20.
64. Mosconi L, Mistur R, Switalski R, Tsui WH, Glodzik L, Li Y, et al. Fdg-pet changes in brain glucose metabolism from normal cognition to pathologically verified Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2009;36:811–22.
65. Kim SH, Seo SW, Yoon DS, Chin J, Lee BH, Cheong H-K, et al. Comparison of neuropsychological and FDG-PET findings between early- versus late-onset mild cognitive impairment: a five-year longitudinal study. *Dement Geriatr Cogn Disord.* 2010;29:213–23.
66. Kljajevic V, Grothe MJ, Ewers M, Teipel S. Distinct pattern of hypometabolism and atrophy in preclinical and predementia Alzheimer's disease. *Neurobiol Aging.* 2014;35:1973–81.
67. Grothe MJ, Silva-Rodríguez J, Moscoco A, Schöll M. FDG-PET is a sensitive imaging biomarker of the neurodegenerative changes that accompany early neurofibrillary tangle pathology. *Alzheimers Dement.* 2022;18:e066508.
68. Gordon BA, Blazey TM, Su Y, Hari-Raj A, Dincer A, Flores S, et al. Spatial patterns of neuroimaging biomarker change in individuals from families with autosomal dominant Alzheimer's disease: a longitudinal study. *Lancet Neurol.* 2018;17:241–50.

69. Ossenkoppele R, Tolboom N, Foster-Dingley JC, Adriaanse SF, Boellaard R, Yaquib M, et al. Longitudinal imaging of Alzheimer pathology using [11C]PIB, [18F]FDDNP and [18F]FDG PET. *Eur J Nucl Med Mol Imaging*. 2012;39:990–1000.
70. Ou Y-N, Xu W, Li J-Q, Guo Y, Cui M, Chen K-L, et al. FDG-PET as an independent biomarker for Alzheimer's biological diagnosis: a longitudinal study. *Alzheimers Res Ther*. 2019;11:57.
71. Gray KR, Wolz R, Heckemann RA, Aljabar P, Hammers A, Rueckert D. Multi-region analysis of longitudinal FDG-PET for the classification of Alzheimer's disease. *Neuroimage*. 2012;60:221–9.
72. Silverman DH, Small GW, Chang CY, Lu CS, Kung De Aburto MA, Chen W, et al. Positron emission tomography in evaluation of dementia: Regional brain metabolism and long-term outcome. *JAMA*. 2001;286:2120–7.
73. Grothe MJ, Moscoso A, Silva-Rodríguez J, Lange C, Nho K, Saykin AJ, et al. Differential diagnosis of amnesic dementia patients based on an FDG-PET signature of autopsy-confirmed LATE-NC. *Alzheimers Dement*. 2023;19:1234–44.
74. Silva-Rodríguez J, Labrador-Espinosa MA, Moscoso A, Schöll M, Mir P, Grothe MJ. Differential effects of tau stage, Lewy body pathology, and substantia nigra degeneration on 18F-FDG PET patterns in clinical Alzheimer disease. *J Nucl Med*. 2023;64:274–80.
75. Wang J, Huang Q, Chen X, You Z, He K, Guo Q, et al. Tau pathology is associated with synaptic density and longitudinal synaptic loss in Alzheimer's disease. *Mol Psychiatry*. 2024;29:2799–809.
76. Vanderlinden G, Ceccarini J, Vande CT, Michiels L, Lemmens R, Triau E, et al. Spatial decrease of synaptic density in amnesic mild cognitive impairment follows the tau build-up pattern. *Mol Psychiatry*. 2022;27:4244–51.
77. Kruggel F, Masaki F, Solodkin A, Alzheimer's Disease Neuroimaging Initiative. Analysis of longitudinal diffusion-weighted images in healthy and pathological aging: an ADNI study. *J Neurosci Methods*. 2017;278:101–15.
78. Hyman BT, Van Hoesen GW, Kromer LJ, Damasio AR. Perforant pathway changes and the memory impairment of Alzheimer's disease. *Ann Neurol*. 1986;20:472–81.
79. Scheltens P, Launer LJ, Barkhof F, Weinstein HC, van Gool WA. Visual assessment of medial temporal lobe atrophy on magnetic resonance imaging: interobserver reliability. *J Neurol*. 1995;242:557–60.
80. Solodkin A, Chen EE, Van Hoesen GW, Heimer L, Shereen A, Kruggel F, et al. In vivo parahippocampal white matter pathology as a biomarker of disease progression to Alzheimer's disease: DTI Biomarkers for Alzheimer's Disease. *J Comp Neurol*. 2013;521:4300–17.
81. Sakaie K, Koenig K, Lerner A, Appleby B, Ogrocki P, Pillai JA, et al. Multi-shell diffusion MRI of the fornix as a biomarker for cognition in Alzheimer's disease. *Magn Reson Imaging*. 2024;109:221–6.
82. Jagust W. Imaging the evolution and pathophysiology of Alzheimer disease. *Nat Rev Neurosci*. 2018;19:687–700.
83. Adams JN, Chappel-Farley MG, Yaros JL, Taylor L, Harris AL, Mikhail A, et al. Functional network structure supports resilience to memory deficits in cognitively normal older adults with amyloid- β pathology. *Sci Rep*. 2023;13:13953.
84. Pignataro A, Meli G, Pagano R, Fontebasso V, Battistella R, Conforto G, et al. Activity-induced amyloid- β oligomers drive compensatory synaptic rearrangements in brain circuits controlling memory of presymptomatic Alzheimer's disease mice. *Biol Psychiatry*. 2019;86:185–95.
85. Vockert N, Machts J, Kleineidam L, Nemali A, Incesoy EI, Bernal J, et al. Cognitive reserve against Alzheimer's pathology is linked to brain activity during memory formation. *Nat Commun*. 2024;15:9815.
86. Elliott ML, Knodt AR, Hariri AR. Striving toward translation: strategies for reliable fMRI measurement. *Trends Cogn Sci*. 2021;25:776–87.
87. Hohenfeld C, Werner CJ, Reetz K. Resting-state connectivity in neurodegenerative disorders: is there potential for an imaging biomarker? *Neuroimage Clin*. 2018;18:849–70.
88. Dauricourt S, de Flores R, Landeau B, Poinsin G, Vanhoutte M, Delcroix N, et al. Longitudinal changes in hippocampal network connectivity in Alzheimer's disease. *Ann Neurol*. 2021;90:391–406.
89. Nyberg L, Salami A, Andersson M, Eriksson J, Kalpouzos G, Kauppi K, et al. Longitudinal evidence for diminished frontal cortex function in aging. *Proc Natl Acad Sci U S A*. 2010;107:22682–6.
90. Enzinger C, Pinter D, Rocca MA, De Luca J, Sastre-Garriga J, Audoin B, et al. Longitudinal fMRI studies: exploring brain plasticity and repair in MS. *Mult Scler*. 2016;22:269–78.
91. Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol*. 2010;9:119–28.
92. Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science*. 1992;256:184–5.
93. Thal DR, Tomé SO. The central role of tau in Alzheimer's disease: from neurofibrillary tangle maturation to the induction of cell death. *Brain Res Bull*. 2022;190:204–17.
94. Jones DT, Knopman DS, Gunter JL, Graff-Radford J, Vemuri P, Boeve BF, et al. Cascading network failure across the Alzheimer's disease spectrum. *Brain*. 2016;139:547–62.
95. Jones DT, Graff-Radford J, Lowe VJ, Wiste HJ, Gunter JL, Senjem ML, et al. Tau, amyloid, and cascading network failure across the Alzheimer's disease spectrum. *Cortex*. 2017;97:143–59.
96. Sanchez JS, Becker JA, Jacobs HIL, Hanseeuw BJ, Jiang S, Schultz AP, et al. The cortical origin and initial spread of medial temporal tauopathy in Alzheimer's disease assessed with positron emission tomography. *Sci Transl Med*. 2021;13:eabc0655.
97. Bilgel M, Wong DF, Moghekar AR, Ferrucci L, Resnick SM, Alzheimer's Disease Neuroimaging Initiative. Causal links among amyloid, tau, and neurodegeneration. *Brain Commun*. 2022;4:fcac193.
98. Mak E, Zhang L, Tan CH, Reilhac A, Shim HY, Wen MOQ, et al. Longitudinal associations between β -amyloid and cortical thickness in mild cognitive impairment. *Brain Commun*. 2023;5:fcad192.
99. Jacobs HIL, Hedden T, Schultz AP, Sepulcre J, Perea RD, Amariglio RE, et al. Structural tract alterations predict downstream tau accumulation in amyloid-positive older individuals. *Nat Neurosci*. 2018;21:424–31.
100. Rieckmann A, Van Dijk KR, Sperling RA, Johnson KA, Buckner RL, Hedden T. Accelerated decline in white matter integrity in clinically normal individuals at risk for Alzheimer's disease. *Neurobiol Aging*. 2016;42:177–88.
101. Harrison TM, La Joie R, Maass A, Baker SL, Swinnerton K, Fenton L, et al. Longitudinal tau accumulation and atrophy in aging and Alzheimer disease. *Ann Neurol*. 2019;85:229–40.
102. Visser D, Verfaillie SCJ, Bosch I, Brouwer I, Tuncel H, Coomans EM, et al. Tau pathology as determinant of changes in atrophy and cerebral blood flow: a multi-modal longitudinal imaging study. *Eur J Nucl Med Mol Imaging*. 2023;50:2409–19.
103. Lattmann-Grefe R, Vockert N, Machts J, Suksangkharn Y, Yakupov R, Schütze H, et al. Dysfunction of the episodic memory network in the Alzheimer's disease cascade. *bioRxiv*. 2024;620237.
104. Gordon BA, McCullough A, Mishra S, Blazey TM, Su Y, Christensen J, et al. Cross-sectional and longitudinal atrophy is preferentially associated with tau rather than amyloid β positron emission tomography pathology. *Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring*. 2018;10:245–52.
105. Pasquini L, Rahmani F, Maleki-Balajoo S, La Joie R, Zarei M, Sorg C, et al. Medial temporal lobe disconnection and hyperexcitability across Alzheimer's disease stages. *Journal of Alzheimer's Disease Reports*. 2019;3:103–12.
106. Gibbons GS, Lee VMY, Trojanowski JQ. Mechanisms of cell-to-cell transmission of pathological tau: a review. *JAMA Neurol*. 2019;76:101–8.
107. Tremblay C, Rahayel S, Pastor-Bernier A, St-Onge F, Vo A, Rheault F, et al. Uncovering atrophy progression pattern and mechanisms in individuals at risk of Alzheimer's disease. *Brain Commun*. 2025;7:099.
108. Leal SL, Landau SM, Bell RK, Jagust WJ. Hippocampal activation is associated with longitudinal amyloid accumulation and cognitive decline. *Elife*. 2017;6:e22978.
109. Fischer L, Molloy EN, Pichet BA, Vockert N, Marquardt J, Pacha PA, et al. Precuneus activity during retrieval is positively associated with amyloid burden in cognitively normal older APOE4 carriers. *J Neurosci*. 2025;6:e1408242024.
110. Moffat G, Zhukovsky P, Coughlan G, Voineskos AN. Unravelling the relationship between amyloid accumulation and brain network function in normal aging and very mild cognitive decline: a longitudinal analysis. *Brain Commun*. 2022;4:fcac282.
111. Palmqvist S, Schöll M, Strandberg O, Mattsson N, Stomrud E, Zetterberg H, et al. Earliest accumulation of β -amyloid occurs within the default-mode network and concurrently affects brain connectivity. *Nat Commun*. 2017;8:1–13.
112. Ziontz J, Harrison TM, Fonseca C, Giorgio J, Han F, Lee J, et al. Connectivity, pathology, and ApoE4 interactions predict longitudinal tau spatial progression and memory. *Hum Brain Mapp*. 2024;45:e70083.
113. Giorgio J, Adams JN, Maass A, Jagust WJ, Breakspear M. Amyloid induced hyperexcitability in default mode network drives medial temporal hyperactivity and early tau accumulation. *Neuron*. 2024;112:676–86.e4.
114. Salami A, Adolfsson R, Andersson M, Blennow K, Lundquist A, Adolfsson AN, et al. Association of APOE ϵ 4 and plasma p-tau181 with preclinical Alzheimer's disease and longitudinal change in hippocampus function. *J Alzheimers Dis*. 2022;85:1309–20.

115. de Calignon A, Polydoro M, Suárez-Calvet M, William C, Adamowicz DH, Kopeikina KJ, et al. Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron*. 2012;73:685–97.
116. Franzmeier N, Neitzel J, Rubinski A, Smith R, Strandberg O, Ossenkoppele R, et al. Functional brain architecture is associated with the rate of tau accumulation in Alzheimer's disease. *Nat Commun*. 2020;11:347.
117. Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci*. 2011;12:723–38.
118. Iadecola C. The pathobiology of vascular dementia. *Neuron*. 2013;80:844–66.
119. Iturria-Medina Y, Sotero RC, Toussaint PJ, Mateos-Pérez JM, Evans AC. Early role of vascular dysregulation on late-onset Alzheimer's disease based on multifactorial data-driven analysis. *Nat Commun*. 2016;7:1–14.
120. Thal DR. The precapillary segment of the blood-brain barrier and its relation to perivascular drainage in Alzheimer's disease and small vessel disease. *Sci World J*. 2009;9:557.
121. Li TR, Li BL, Xu XR, Zhong J, Wang TS, Liu FQ, et al. Association of white matter hyperintensities with cognitive decline and neurodegeneration. *Front Aging Neurosci*. 2024;16:1412735.
122. Luo J, Ma Y, Agboola FJ, Grant E, Morris JC, McDade E, et al. Longitudinal relationships of white matter hyperintensities and Alzheimer disease biomarkers across the adult life span. *Neurology*. 2023;101:e164–77.
123. Fíford CM, Manning EN, Bartlett JW, Cash DM, Malone IB, Ridgway GR, et al. White matter hyperintensities are associated with disproportionate progressive hippocampal atrophy. *Hippocampus*. 2017;27:249–62.
124. Bernal J, Menze I, Yakupov R, Peters O, Hellmann-Regen J, Freiesleben SD, et al. Longitudinal evidence for a mutually reinforcing relationship between white matter hyperintensities and cortical thickness in cognitively unimpaired older adults. *Alzheimers Res Ther*. 2024;16:1–14.
125. Tamagno E, Guglielmotto M, Vasciaveo V, Tabaton M. Oxidative stress and beta amyloid in Alzheimer's disease. Which comes first: the chicken or the egg? *Antioxidants*. 2021;10:1479.
126. Cha W-J, Yi D, Ahn H, Byun MS, Chang YY, Choi J-M, et al. Association between brain amyloid deposition and longitudinal changes of white matter hyperintensities. *Alzheimers Res Ther*. 2024;16:1–9.
127. Moscoso A, Rey-Bretal D, Silva-Rodríguez J, Aldrey JM, Cortés J, Pías-Peleiteiro J, et al. White matter hyperintensities are associated with subthreshold amyloid accumulation. *Neuroimage*. 2020;218:116944.
128. Li Y, Kalpouzos G, Laukka EJ, Dekhtyar S, Bäckman L, Fratiglioni L, et al. Progression of neuroimaging markers of cerebral small vessel disease in older adults: a 6-year follow-up study. *Neurobiol Aging*. 2022;112:204–11.
129. Menze I, Bernal J, Kaya P, Aki Ç, Pfister M, Geisendörfer J, et al. Perivascular space enlargement accelerates in ageing and Alzheimer's disease pathology: evidence from a three-year longitudinal multicentre study. *Alzheimers Res Ther*. 2024;16:242.
130. Fan Z, Okello AA, Brooks DJ, Edison P. Longitudinal influence of microglial activation and amyloid on neuronal function in Alzheimer's disease. *Brain*. 2015;138:3685–98.
131. Edison P, Ahmed I, Fan Z, Hinz R, Gelosa G, Ray Chaudhuri K, et al. Microglia, amyloid, and glucose metabolism in Parkinson's disease with and without dementia. *Neuropsychopharmacology*. 2013;38:938–49.
132. Sanchez-Guajardo V, Barnum CJ, Tansey MG, Romero-Ramos M. Neuroimmunological processes in Parkinson's disease and their relation to α -synuclein: microglia as the referee between neuronal processes and peripheral immunity. *ASN Neuro*. 2013;5:113–39.
133. Marlatt MW, Bauer J, Aronica E, van Haastert ES, Hoozemans JJM, Joels M, et al. Proliferation in the Alzheimer hippocampus is due to microglia, not astroglia, and occurs at sites of amyloid deposition. *Neural Plast*. 2014;2014:693851.
134. Heneka MT, van der Flier WM, Jessen F, Hoozemans J, Thal DR, Boche D, et al. Neuroinflammation in Alzheimer disease. *Nat Rev Immunol*. 2024;25:321–52. <https://doi.org/10.1038/s41577-024-01104-7>.
135. Ismail R, Parbo P, Madsen LS, Hansen AK, Hansen KV, Schaldemose JL, et al. The relationships between neuroinflammation, beta-amyloid and tau deposition in Alzheimer's disease: a longitudinal PET study. *J Neuroinflammation*. 2020;17:151.
136. Bradburn S, Murgatroyd C, Ray N. Neuroinflammation in mild cognitive impairment and Alzheimer's disease: a meta-analysis. *Ageing Res Rev*. 2019;50:1–8.
137. Finze A, Biechele G, Rauchmann B-S, Franzmeier N, Palleis C, Katzdobler S, et al. Individual regional associations between $A\beta$, tau- and neurodegeneration (ATN) with microglial activation in patients with primary and secondary tauopathies. *Mol Psychiatry*. 2023;28:4438–50.
138. Appleton J, Finn Q, Zanotti-Fregonara P, Yu M, Faridar A, Nakawah MO, et al. Brain inflammation co-localizes highly with tau in mild cognitive impairment due to early-onset Alzheimer's disease. *Brain*. 2025;148:119–32.
139. Rossano SM, Johnson AS, Smith A, Ziaggi G, Roetman A, Guzman D, et al. Microglia measured by TSPO PET are associated with Alzheimer's disease pathology and mediate key steps in a disease progression model. *Alzheimers Dement*. 2024;20:2397.
140. Zou J, Tao S, Johnson A, Tomljanovic Z, Polly K, Klein J, et al. Microglial activation, but not tau pathology, is independently associated with amyloid positivity and memory impairment. *Neurobiol Aging*. 2020;85:11–21.
141. Snellman A, Ekblad LL, Tuisku J, Koivumäki M, Ashton NJ, Lantero-Rodriguez J, et al. APOE $\epsilon 4$ gene dose effect on imaging and blood biomarkers of neuroinflammation and beta-amyloid in cognitively unimpaired elderly. *Alzheimers Res Ther*. 2023;15:1–15.
142. Kreisler WC, Lyoo CH, Liow J-S, Wei M, Snow J, Page E, et al. (11)C-PBR28 binding to translocator protein increases with progression of Alzheimer's disease. *Neurobiol Aging*. 2016;44:53–61.
143. Villemagne VL, Pike KE, Chételat G, Ellis KA, Mulligan RS, Bourgeat P, et al. Longitudinal assessment of $A\beta$ and cognition in aging and Alzheimer disease. *Ann Neurol*. 2011;69:181–92.
144. Fonseca CS, Baker SL, Dobyns L, Janabi M, Jagust WJ, Harrison TM. Tau accumulation and atrophy predict amyloid independent cognitive decline in aging. *Alzheimer's & Dementia*. 2024;20:2526–37.
145. Chen X, Juarez A, Mason S, Kobayashi S, Baker SL, Harrison TM, et al. Longitudinal relationships between $A\beta$ and tau to executive function and memory in cognitively normal older adults. *Neurobiol Aging*. 2025;145:32–41.
146. Farrell ME, Papp KV, Buckley RF, Jacobs HL, Schultz AP, Properzi MJ, et al. Association of emerging β -amyloid and tau pathology with early cognitive changes in clinically normal older adults. *Neurology*. 2022;98:e1512–24.
147. Farrell ME, Thibault EG, Alex Becker J, Price JC, Healy BC, Hanseeuw BJ, et al. Spatial extent as a sensitive amyloid-PET metric in preclinical Alzheimer's disease. *Alzheimer's & Dementia*. 2024;20:5434–49.
148. Boccalini C, Ribaldi F, Hristovska I, Arnone A, Peretti DE, Mu L, et al. The impact of tau deposition and hypometabolism on cognitive impairment and longitudinal cognitive decline. *Alzheimer's & Dementia*. 2024;20:221–33.
149. Malpetti M, Kievit RA, Passamonti L, Jones PS, Tsvetanov KA, Rittman T, et al. Microglial activation and tau burden predict cognitive decline in Alzheimer's disease. *Brain*. 2020;143:1588–602.
150. Ottroy J, Niemantsverdriet E, Verhaeghe J, De Roeck E, Struyfs H, Somers C, et al. Association of short-term cognitive decline and MCI-to-AD dementia conversion with CSF, MRI, amyloid- and F-FDG-PET imaging. *Neuroimage Clin*. 2019;22:101771.
151. Hou M, Bergamino M, de Chastelaine M, Sambamoorthy S, Rugg MD. Free water-corrected fractional anisotropy of the fornix and parahippocampal cingulum predicts longitudinal memory change in cognitively healthy older adults. *Neurobiol Aging*. 2024;142:17–26.
152. Song Z, Farrell ME, Chen X, Park DC. Longitudinal accrual of neocortical amyloid burden is associated with microstructural changes of the fornix in cognitively normal adults. *Neurobiol Aging*. 2018;68:114–22.
153. Ferreira D, Verhagen C, Hernández-Cabrera JA, Cavallin L, Guo C-J, Ekman U, et al. Distinct subtypes of Alzheimer's disease based on patterns of brain atrophy: longitudinal trajectories and clinical applications. *Sci Rep*. 2017;7:46263.
154. Gorbach T, Pudas S, Lundquist A, Orädd G, Josefsson M, Salami A, et al. Longitudinal association between hippocampal atrophy and episodic-memory decline. *Neurobiol Aging*. 2017;51:167–76.
155. Shokouhi S, Claassen D, Kang H, Ding Z, Rogers B, Mishra A, et al. Longitudinal progression of cognitive decline correlates with changes in the spatial pattern of brain 18F-FDG PET. *J Nucl Med*. 2013;54:1564–9.
156. Groot C, Risacher SL, Chen JQA, Dicks E, Saykin AJ, CI MD, et al. Differential trajectories of hypometabolism across cognitively-defined Alzheimer's disease subgroups. *Neuroimage Clin*. 2021;31:102725.
157. Staffaroni AM, Cobigo Y, Elahi FM, Casaletto KB, Walters SM, Wolf A, et al. A longitudinal characterization of perfusion in the aging brain and associations with cognition and neural structure. *Hum Brain Mapp*. 2019;40:3522–33.
158. O'Brien JL, O'Keefe KM, LaViolette PS, DeLuca AN, Blacker D, Dickerson BC, et al. Longitudinal fMRI in elderly reveals loss of hippocampal activation with clinical decline. *Neurology*. 2010;74:1969–76.
159. Huijbers W, Mormino EC, Schultz AP, Wigman S, Ward AM, Larvie M, et al. Amyloid- β deposition in mild cognitive impairment is associated with increased hippocampal activity, atrophy and clinical progression. *Brain*. 2015;138:1023–35.

160. Staffaroni AM, Brown JA, Casaletto KB, Elahi FM, Deng J, Neuhaus J, et al. The longitudinal trajectory of default mode network connectivity in healthy older adults varies as a function of age and is associated with changes in episodic memory and processing speed. *J Neurosci*. 2018;38:2809–17.
161. Fischer L, Adams JN, Molloy EN, Vockert N, Tremblay-Mercier J, Remz J, et al. Differential effects of aging, Alzheimer's pathology, and APOE4 on longitudinal functional connectivity and episodic memory in older adults. *Alzheimers Res Ther*. 2025;17:1–20.
162. Al-Janabi OM, Bauer CE, Goldstein LB, Murphy RR, Bahrani AA, Smith CD, et al. White matter hyperintensity regression: comparison of brain atrophy and cognitive profiles with progression and stable groups. *Brain Sci*. 2019;9:170.
163. Young AL, Marinescu RV, Oxtoby NP, Bocchetta M, Yong K, Firth NC, et al. Uncovering the heterogeneity and temporal complexity of neurodegenerative diseases with subtype and stage inference. *Nat Commun*. 2018;9:4273.
164. Young AL, Oxtoby NP, Garbarino S, Fox NC, Barkhof F, Schott JM, et al. Data-driven modelling of neurodegenerative disease progression: thinking outside the black box. *Nat Rev Neurosci*. 2024;25:111–30.
165. Smith R, Capotosti F, Schain M, Ohlsson T, Vokali E, Molette J, et al. The α -synuclein PET tracer [18F] ACI-12589 distinguishes multiple system atrophy from other neurodegenerative diseases. *Nat Commun*. 2023;14:6750.
166. Franzmeier N, Roemer-Cassiano SN, Bernhardt AM, Dehsarvi A, Dewenter A, Steward A, et al. Alpha synuclein co-pathology is associated with accelerated amyloid-driven tau accumulation in Alzheimer's disease. *Mol Neurodegener*. 2025;20:1–15.
167. Betthauser TJ. In vitro evidence for a nonselective 4R tau PET tracer. *Mol Psychiatry*. 2023;28:1398–9.
168. Bavarsad MS, Grinberg LT. SV2A PET imaging in human neurodegenerative diseases. *Front Aging Neurosci*. 2024;16:1380561.
169. Cai Z, Li S, Matuskey D, Nabulsi N, Huang Y. PET imaging of synaptic density: a new tool for investigation of neuropsychiatric diseases. *Neurosci Lett*. 2019;691:44–50.
170. Cortes-Canteli M, Iadecola C. Alzheimer's disease and vascular aging: JACC focus seminar. *J Am Coll Cardiol*. 2020;75:942–51.
171. Perosa V, Oltmer J, Munting LP, Freeze WM, Auger CA, Scherlek AA, et al. Perivascular space dilation is associated with vascular amyloid- β accumulation in the overlying cortex. *Acta Neuropathol*. 2022;143:331–48.
172. de Godoy LL, Alves CAPF, Saavedra JSM, Studart-Neto A, Nitrini R, da Costa Leite C, et al. Understanding brain resilience in superagers: a systematic review. *Neuroradiology*. 2021;63:663–83.
173. Bocancea DI, Svenningsson AL, van Loenhoud AC, Groot C, Barkhof F, Strandberg O, et al. Determinants of cognitive and brain resilience to tau pathology: a longitudinal analysis. *Brain*. 2023;146:3719–34.
174. Zhang M, Ganz AB, Rohde S, Rozemuller AJM, Bank NB, Reinders MJT, et al. Resilience and resistance to the accumulation of amyloid plaques and neurofibrillary tangles in centenarians: an age-continuous perspective. *Alzheimers Dement*. 2023;19:2831–41.
175. Weiner MW, Veitch DP, Miller MJ, Aisen PS, Alcala B, Beckett LA, et al. Increasing participant diversity in AD research: plans for digital screening, blood testing, and a community-engaged approach in the Alzheimer's Disease Neuroimaging Initiative 4. *Alzheimers Dement*. 2023;19:307–17.
176. Meeker KL, Wisch JK, Hudson D, Coble D, Xiong C, Babulal GM, et al. Socio-economic status mediates racial differences seen using the AT(N) framework. *Ann Neurol*. 2021;89:254–65.
177. Lachner C, Craver EC, Babulal GM, Lucas JA, Ferman TJ, White RO, et al. Disparate dementia risk factors are associated with cognitive impairment and rates of decline in African Americans. *Ann Neurol*. 2024;95:518–29.
178. Arenaza-Urquijo EM, Boyle R, Casaletto K, Anstey KJ, Vila-Castelar C, Colverson A, et al. Sex and gender differences in cognitive resilience to aging and Alzheimer's disease. *Alzheimers Dement*. 2024;20:5695–719.
179. Livingston G, Huntley J, Liu KY, Costafreda SG, Selbæk G, Alladi S, et al. Dementia prevention, intervention, and care: 2024 report of the Lancet standing Commission. *Lancet*. 2024;404:572–628.
180. Cummings J, Osse AML, Cammann D, Powell J, Chen J. Anti-amyloid monoclonal antibodies for the treatment of Alzheimer's disease. *BioDrugs*. 2024;38:5–22.
181. Spires-Jones TL, Attems J, Thal DR. Interactions of pathological proteins in neurodegenerative diseases. *Acta Neuropathol*. 2017;134:187–205.
182. Robinson JL, Xie SX, Baer DR, Suh E, Van Deerlin VM, Loh NJ, et al. Pathological combinations in neurodegenerative disease are heterogeneous and disease-associated. *Brain*. 2023;146:2557–69.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.